

ORIGINAL

LAW OFFICES OF

WILLIAMS LOVE O'LEARY & POWERS, P.C.

MICHAEL L. WILLIAMS
mwilliams@wdolaw.com

LINDA C. LOVE
llove@wdolaw.com

SUITE 450
9755 SW BARNES ROAD
PORTLAND, OREGON 97225-6681

TELEPHONE 503/295-2924
FAX NUMBER 503/295-3720
TOLL FREE 800/842-1595
WWW.WDOLAW.COM

LESLIE W. O'LEARY
loleary@wdolaw.com

THOMAS B. POWERS
tpowers@wdolaw.com

August 31, 2007

VIA UPS DELIVERY



Clerk
United States Court of Federal Claims
717 Madison Place, NW
Washington, D.C. 20005

Re: In Re: Claims for Vaccine Injuries Resulting in Autism Spectrum Disorder, or a Similar
Neurodevelopmental Disorder v. Secretary of Health And Human Services
Autism Master File
Our File No. 054500 - Omnibus Autism Proceeding

Dear Clerk of Court:

Enclosed for filing in the Autism Master File is the original and two copies of the report of petitioners' expert Dr. H. Vasken Aposhian, PhD, including his current curriculum vitae and a bibliography of literature in support of his expert opinion. This report is submitted in support of petitioners' theory that exposure to the mercury contained in certain pediatric vaccines was a substantial contributing cause of some or all of the injuries at issue in the Omnibus Autism Proceeding. Petitioners' anticipate relying on this testimony in hearings on "test cases" currently scheduled for May 2008.

Very truly yours,

A handwritten signature in black ink, appearing to read "T. B. Powers". The signature is fluid and cursive, with a long, sweeping tail.

Thomas B. Powers
Attorney at Law

Enclosures

cc: John Fabry, Esq., Williams Bailey Law Firm, LLP (via e-mail)
Vincent J. Matanoski, Esq., U.S. Department of Justice (via UPS delivery)

CERTIFICATE OF SERVICE

I hereby certify that on August 31, 2007, I served the foregoing **Expert Report from Dr.**

H. Vasken Aposhian, PhD on the following individual(s):

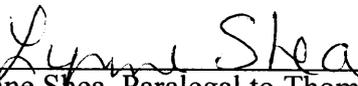
John Fabry, Esq.
Williams Kherkher Hart Boundas, LLP
8441 Gulf Freeway, Suite 600
Houston, TX 77017-5001

Vincent Matanoski, Esq.
Mark Raby, Esq.
US Department of Justice
Torts Branch, Civil Division
1425 New York Avenue NW
Suite 3100
Washington DC, 20005

By United Parcel Service, next day delivery.

Petitioners specifically authorize the Court and the Office of Special Masters to post this document, and any attachments or exhibits thereto, on the Court/OSM website, expressly waiving any confidentiality as to the contents of these materials. Petitioners expressly wish to publicly disclose this filing in any other forum designated by the Court or the OSM.

WILLIAMS LOVE O'LEARY & POWERS, P.C.



Lynne Shea, Paralegal to Thomas B. Powers
Of Attorneys for Petitioners' Steering Committee



**Thimerosal and its biotransformant ethylmercury cause injury to the
developing brains of human embryos and young children.**

H. Vasken Aposhian, PhD

Professor of Molecular and Cellular Biology

Professor of Pharmacology

LSSB Rm 444

The University of Arizona

PO BOX 210106

Tucson, AZ 85721-0106

(520) 621-7565

FAX (520) 621-3709

August 30, 2007

Outline

1. Introduction
2. Brief overview of the chemical forms of mercury
3. There is evidence for genetic differences (hypersusceptibility) in humans for response to mercury exposure.
4. There are specific windows for brain development in the human embryo, fetus and infant.
 - a. The specific developmental windows of thalidomide for the human embryo and the lessons learned from thalidomide are guides for understanding such effects of other agents.
 - b. Recent evidence demonstrates that terbutaline (a drug used to treat premature labor) is a developmental neurotoxicant and may cause brain damage in the infant.
5. Thimerosal containing vaccines will result in toxic levels of mercury in the brain.
6. Conclusion: Thimerosal and its biotransformant ethylmercury cause injury to the developing brains of human embryos and young children by interfering with brain development during highly specific windows of brain development. These windows of susceptibility may occur during the prenatal and/or postnatal periods. These windows are vulnerable and open at very specific times. Thus, thimerosal induces symptoms of autism spectrum disorders in some subsets of children.

There is growing importance of neuroinflammatory events in the development of autism. All of this indicates the need for closer examination and better correlations of past research on the genetics and environmental factors causing autism. One of these environmental factors is surely thimerosal..

Professional Qualifications and Experience

My curriculum vitae is appended hereto as Appendix "A". As it shows, I am Professor of Molecular and Cellular Biology, Faculty of Science, and Professor of Pharmacology, College of Medicine, at The University of Arizona, Tucson, Arizona. I received my undergraduate education (Sc. B. in Chemistry) at Brown University and was awarded a Ph.D. in Physiological Chemistry from The University of Rochester. I have been a member of the basic science departments of a number of medical schools, including Vanderbilt, Stanford, and Tufts. Early in my professional education and training, I spent three years doing research as a National Institute of Health Senior Postdoctoral Fellow under the tutelage of Nobel Laureate Dr. Arthur Kornberg, Professor of Biological Chemistry, College of Medicine, Stanford University. I was brought to The University of Arizona in 1975 as Department Head to modernize the biology departments. I have been elected to a number of professional societies such as the Society of Toxicology, The American Society of Biological Chemistry and Molecular Biology, The American Association for the Advancement of Science, The American Society of Microbiology and others.

I have published numerous peer-reviewed research papers in first-class international scientific journals dealing with my research on DNA, gene transfer, treatments for and mechanisms of the toxicology of lead, arsenic and mercury and other heavy metals. My laboratory is still active in research dealing with mercury, arsenic as well as chelating agents. A recent article (Aposhian and Aposhian, 2006) from my laboratory was the most downloaded article in 2006 of the preeminent toxicology journal *Chemical Research in Toxicology*.

I have been on numerous scientific committees of the federal government including the White House Conference on Mercury; the National Academy of Science/National Research Committees that wrote the critical monograph *Toxicological Effects of Methyl Mercury* and the

monograph on *Arsenic in Drinking Water*. I wrote the toxicology chapter of the former monograph. I have been a member of numerous Committees of the National Institutes of Health, the Environmental Protection Agency, the National Science Foundation and a number of Foundations. I have studied mercury and/or arsenic in people in Chile, Inner Mongolia, Mexico, China, and Romania. The visits to these foreign countries to do the studies for research purposes required scientific knowledge, tact, and unusual communication skills. I am still active in teaching undergraduate human toxicology.

My research has been supported since 1955 by one or more grants from the National Institute of Health, the U. S. Army, the National Science Foundation, The Wallace Research Foundation, The Autism Research Institute and other foundations. I have been a consultant to a number of multi-national pharmaceutical firms. I have given, by invitation, innumerable presentations on the results of my scientific research at many universities, pharmaceutical companies, and national and international government organizations. I have been a National Academy of Science Visiting Fellow to the former USSR and an official guest in the Soviet Union, China and Mexico to deal with mercury and arsenic health problems. I have been very fortunate in life being paid to do what I enjoy doing.

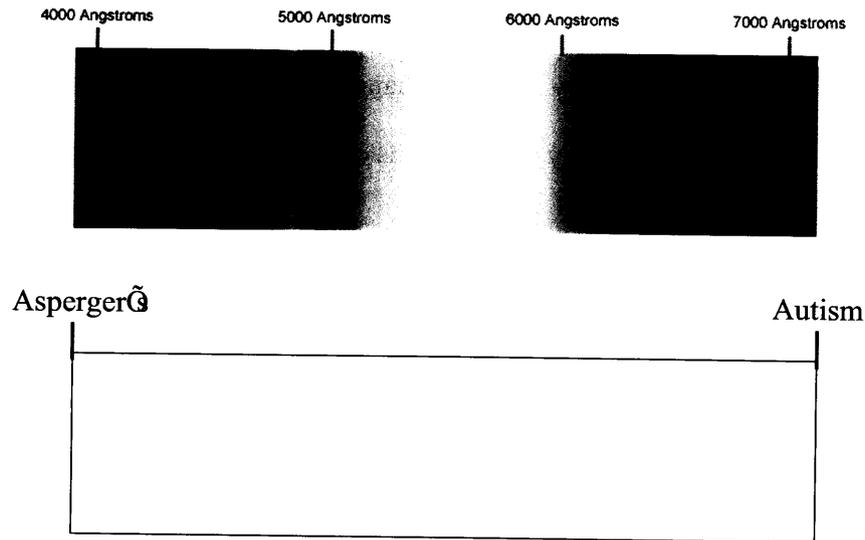
1- Introduction

The purpose of this report is to give the Court information as to the possibility that some cases of autism are caused by a direct action of thimerosal and/or its metabolite ethyl mercury on the developing brain/central nervous system. This happens during certain vulnerable windows of the developmental processes of the very young brains of embryos and/ or children.

Autism spectrum disorders (ASD) are composed of a broad spectrum of neurodevelopmental disorders known as pervasive developmental disorders. They occur in childhood and are characterized by impairments in social interaction, verbal and nonverbal communication and restricted and repetitive stereotyped behaviors. The etiology of ASD is largely unknown. However, genetic, environmental, immunological, and neurological factors are believed to have a role in its development of this disorder. The onset of ASD usually occurs in the first 36 months of childhood. The CDC has recently stated that the prevalence of ASD is as high as 1 per 150 children.

One of the problems of trying to understand the etiology of ASD is that at one end of the spectrum is severe autism at the other end is Asperger's Disorder (Figure 1) . When a chemist looks at or studies a spectrum, he looks at the individual defined bands. Unfortunately, this has not been done with ASD. It appears that very little has been done to characterize each of the individual disorders or each band that characterize ASD.

Figure 1. The visible spectrum and autism spectrum disorders



Where are the bands of ASD?

Another point that is pertinent to make in this introduction is that one needs to realize that until recently the once accepted toxicology axiom that the dose determines the poison is no longer 100% correct. Many other factors are involved in vulnerability of a person to poisons. Some of them are listed below

- i) Antibiotics being used when mercury exposure occurs can inhibit mercury excretion and thus potentially increase its toxicity. (Rowland et al., 1984)
- ii) A combination of a genetic predisposition and a stress (e.g. a fever) may increase the impact of the stress-causing agent. (Morton et al., 1991)

iii) The environment is of course important: The more mercury in the air the greater the incidence of autism. (Palmer et al., 2006)

iv) Diet can make a child more vulnerable to autism. Glutamine is low in autistic children. Glutamine is a precursor of glutathione. Glutathione is a major factor and is necessary for metal transport in the body and detoxification processes. (Nordberg et al., 2007).

2. Brief overview of the chemical forms of mercury

Although the toxicology of the various chemical forms (sometimes called species) have been presented in the Federal Vaccine Court Cedillo proceedings it is pertinent to extend the previously submitted information.

Mercuric mercury (Hg^{+2}) is known to bind strongly to sulfhydryl groups, for example those of proteins and glutathione (GSH). The ultimate toxin for organic mercury compounds such as methyl mercury (MeHg) or ethyl mercury (EtHg), however, is still unknown. It could be MeHg (EtHg), per se, mercuric mercury (Hg^{++}) or a combination of both Hg^{++} and the organic mercury compound. In addition it is not clear as to whether MeHg or EtHg causes cell death by binding to sulfhydryl groups as does mercuric mercury. MeHg, for example, causes many different deviations from the usual cause of cell death. These include impaired glycolysis, impaired nucleic acid biosynthesis, impaired aerobic respiration, impaired protein synthesis (Cheung and Verity, 1985), and impaired neurotransmitter release (Atchinson and Hare, 1994).

3. There is evidence for genetic differences (hypersusceptibility) in response to mercury exposure.

The brain and central nervous system (CNS) not only develop in utero but continue to develop until at least puberty. The stages of CNS development are often described as windows of development and the windows of many toxic chemicals are known.

It has been known for some time that heavy metals disrupt the normal metabolism for example, heme metabolism (Heyer et al. 2006). This is usually expressed by a change in the urinary porphyrin profile. Urinary porphyrins of dentists with low-level mercury exposure were analyzed. The expected urinary porphyrin profile was found in 85% of the dentists. But 15% had an atypical porphyrinogenic response. The latter was due to polymorphism in the human gene that modifies the effect of Hg on a biological process. This human gene is the coproporphyrinogen oxidase gene. (Woods et al., 2005). The result of the polymorphism is the inhibition of an enzyme and the occurrence of a new porphyrin in the urine. Although this work was published over two years ago, many toxicologists do not cite it in their published review articles because they are not cognizant or familiar with the importance of genetics and genetic toxicology as an important area of human toxicology. Research into the genetics of mercury toxicity is just beginning.

The Woods et al., (2005) work has been followed by a paper from France (Nataf et al., 2006) involving 115 autistic and 119 control children. They showed that urine coproporphyrin levels were elevated in autistic children. Urinary precoproporphyrin, an indicator of heavy metal toxicity was also elevated in autism. When subgroups of these children were given the chelating agent DMSA, there was a reduction in the mean urinary levels of precoproporphyrin and coproporphyrin for the autistic children.

Another example of hypersusceptibility to mercury is Pink Disease. During the years 1850 through 1955, a devastating syndrome called Pink Disease or acrodynia affected many

infant children (see Dally, 1997, for an excellent review). It is now believed that a teething powder that contained mercurous salts caused this illness.

While review articles exist about “Mercury-the metal of mystery” (e.g. Clarkson, 2002; Clarkson et al., 2003; Clarkson and Magos, 2006; and Counter and Buchanan, 2004), they must be viewed cautiously as current scientific investigations may render some of their conclusions false, inaccurate or outdated. After all, one definition of science is the search for the truth. Scientific investigation is always ongoing and cannot be stagnant. Science always progresses and the resultant knowledge changes with time. This concern about inaccurate and outdated conclusions has also been voiced by Mutter et al., (2007)

It is appropriate to distinguish between two very similar concepts, heritability and genetic causation. Heritability involves similar traits seen between parents and offspring. For example among Europeans the wearing of skirts is a very strong heritability (it occurs only among women, except for an occasional Scot). The wearing of a skirt thus is related to having two X chromosomes but the trait is not genetically determined. On the other hand the number of fingers on a human hand is completely determined by genetics. When there is a deviation from the normal five-fingered hand it is caused by defects in development. Thalidomide, a potent teratogen, causing severe birth defects is an example of this and is not heritable. A better discussion of this can be found in Vineis and Kriebal (2006).

4. There are specific windows for brain development in the human embryo, fetus and infant.

a. The specific developmental windows of thalidomide for the human embryo and the lessons learned from thalidomide are guides for understanding such effects of other agents.

The concept that the toxicity of a chemical depends on when the exposure to that toxin occurred is well demonstrated by thalidomide. This agent was developed and used in Germany and many other countries as a sedative. In the late 1950 to early 1960's a large number of malformed children were born in Germany and Austria. Some were born with phocomelia (a developmental anomaly in which the proximal portion of a limb or the complete limb is absent, - the hands or feet are attached directly to the trunk of the body), some with amelia (the absence of hands and limbs), and/or other developmental defects. There were at least 6000 cases of thalidomide-caused live births with birth defects. By excellent medical investigations, it was demonstrated that these birth defects were caused by women using thalidomide as a sedative at some time during their pregnancy. What soon became apparent was that the type of defect depended on when during the gestation period the thalidomide had been taken. See figures taken from Miller and Stromland (1999) below. Thalidomide was found to be teratogenic between 20 to 36 days after fertilization (34-50 days after the last menstrual cycle). The windows for phocomelia and amelia have been narrowed down to specific periods of 1 to 2 days. Other windows for other thalidomide-caused developmental defects were also discovered.

Since then developmental windows in which other drugs and chemicals exert their effects have been discovered. Thalidomide was the initial reason for the development of a new area of

scientific investigation known as teratogens that is medically defined as agents causing monster formation.

Not all children whose mothers took thalidomide during their pregnancies were born with deformities. For teratogenicity to occur, the mother must be predisposed to having the drug act in that manner and the embryo must be predisposed (genetic!) to its action. In addition, thalidomide differs from some other teratogens in that the **time of administration of the drug appears to be more important a factor than the dosage** (Miller and Stromland, 1999)

Figure 2. Miller and Stromland (1999) *Image unavailable at time of filing*

Figure 3. Miller and Stromland (1999)

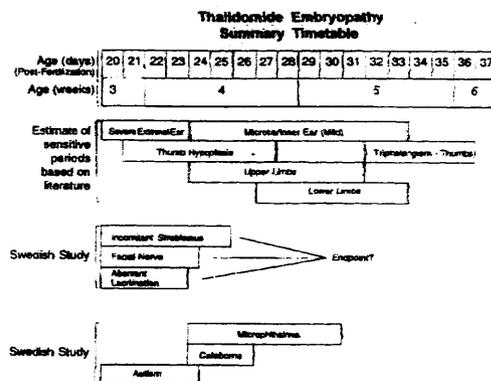
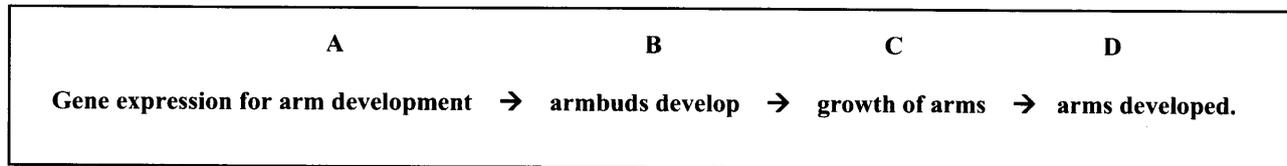


Fig. 7. Summary estimates of sensitive periods for ophthalmic malformations based on associated anomalies manifested by most patients (see Fig. 6 for Duane syndrome, the most frequent incomitant strabismus). The other clinical forms of incomitant strabismus had similar associated anomalies. These sensitive time blocks for the development of eye anomalies and autism are estimations and they may be shorter or slightly longer than indicated. The thalidomide timetable for the ear, thumb, and limb anomalies was derived from the literature (Nowack, '65; Lenz, '66; Kida, '87).

Thus, it is very reasonable and likely that thimerosal/ethylmercury (like thalidomide and many other agents) have specific windows of brain toxicity. Mercury (thimerosal/ethylmercury) containing vaccines (MCV's) given on typical US schedules in the 1990's (which called for 187.5 micrograms of mercury to be injected into virtually every child by six months of age and

which started on the day of birth) likely contributed to the development of Autism Spectrum Disorders in a susceptible small subset of those children. Also it must be remembered that windows of vulnerability vary within a very small time period because everything biological has a certain degree of variability. Thus the reason why some children got autism while other did not was they were not all vaccinated at precisely the same time. For example some received their 2 month vaccination precisely on the 60th day of their life; others maybe on their 57 th day of life; others on their 64th day of life and so on. The perturbations would be worthy of a mathematician. It is also very important to realize that the effects of thalidomide teratogenicity are delayed. For example



Another way of looking at this pathway is A>B>C>D.

The arm develops along the A to D pathway. Thalidomide acts during A, B or C not at D.

So most agents acting on such windows have a delayed response. Thalidomide causes phocomelia but it acts before the actual arms have developed. In other words there is a **delay** in the manifestations to its exposure. This is true of most developmental toxins.

b. Recent evidence indicates that terbutaline (a drug used for premature labor) is a developmental neurotoxicant and may cause brain damage in the infant (Rhodes et al, 2004).

Terbutaline is a β_2 -adenoceptor agonist that crosses the placenta and enters the fetal brain to stimulate fetal β_2 -adenoceptors that control neural cell development. This drug causes

biochemical changes and structural damage in the immature brain during critical times of such development (Rhodes et al., 2004). These effects strongly indicate that fetal terbutaline exposure causes a higher incidence of cognitive and neuropsychiatric disorders for the children born of women given terbutaline treatment for preterm labor.

These two drugs, thalidomide and terbutaline, demonstrate in the former case a well established and long-accepted developmental toxicity of a drug given during the time and windows of fetal brain development. For the latter drug, such information is more recent but just as important. All this information is relevant as to how thimerosal/ethylmercury can act to damage the development of the brain in embryos and young children. Furthermore, Zerrate et al. (2007) have developed a rat model in which terbutaline given at very specific times in the window of brain development causes overstimulation of β_2 -adrenoceptor resulting in microglial activation associated with innate neuroinflammatory pathways and behavioral abnormalities similar to those seen for autism.

5- The Brain and Mercury

How much mercury gets into an infant's brain after an injection of thimerosal containing vaccine? We can make calculations as follows based on the papers of Burbacher et al., (2005) and Pichichero et al., (2002).

Burbacher et al., (2005) demonstrated that the brain to blood concentration ratio was 3.5 ± 0.5 in infant monkeys receiving thimerosal.

Pichichero et al., (2002) using **nonautistic** children showed that

	<u>2-month-old children</u>	<u>6-month-old children</u>
Blood Hg concentration range	4.50 to 20.6 nanoMolar	2.85 to 6.9 nanoMolar
Mean	8.20 nanoMolar ± 4.85 SD	5.15 nanoMolar ± 1.20 SD

From the above and the Burbacher derived brain to blood mercury ratio in infant monkeys, the following can be calculated for human infants after receiving a thimerosal-vaccine injection

	<u>2-month-old children</u>	<u>6-month-old children</u>
Brain Hg range	15.75 to 72.1 nanoMolar	9.98 to 24.15 nanoMolar
Mean	28.7 nanoMolar	18.0 nanoMolar

Controls with no thimerosal had no quantifiable Hg except for one of 15 control infants.

It is reasonable than that 2-month-old infants have as much as 72.1 nanoMolar brain Hg. It needs to be remembered, however, that the above data are based on investigations of normal, not autistic children. If there is a subset of autistic children with a mercury efflux disorder, they would be expected to have even greater concentrations of mercury in their tissues. Also when we are able to calculate back to the time of the peak of the bolus injections of thimerosal containing vaccines, the concentration of brain mercury would be expected to be even greater.

Neuroinflammation and autism. There is increasing evidence for neuroinflammatory events being involved in the development of autism. It should be kept in mind that the term “neuroinflammation” was not even found in the scientific literature before 1995. This thinking is very recent and is just the ”tip of the iceberg” as to what we expect to learn about immunity, neuroglia and neuroinflammation in autism. The presence of neuroglial and innate neuroimmune system activation in brain tissue and cerebrospinal fluid of autistics supports the theory that neuroimmune abnormalities occur in their brain (Pardo et al. (2005).

Connors et al. (2005) from Johns Hopkins Medical Center have investigated certain activities and genetic polymorphisms in autism using dizygotic twins. They concluded that prenatal overstimulation of the β_2 -adrenergic receptor by terbutaline or by the increased signaling caused by polymorphisms in the genes of this receptor having diminished desensitization can influence the cellular responses and developmental programs of the fetal brain leading to autism.

Courchesne et al., (2005) from the University of California, San Diego have argued that in autism, higher order functions appear to fail to develop normally because frontal, cerebellar and temporal cellular and growth pathologies occur prior to and during the critical period, much of it immediately after birth, when these higher order brain areas first begin to form their neurological circuitry. They go on to state that “this altered circuitry impairs the essential role of frontal cortex in integrating information from diverse functional systems (emotional, sensory, autonomic, memory, etc) and providing context-based and goal-directed feedback to lower level systems.”

With the above in mind it is not difficult to hypothesize the effects of thimerosal and/or ethylmercury on the developing brain. However, although the purpose of this report is to examine the likelihood that thimerosal and/or ethylmercury causes damage to the developing brain of a subpopulation of hypersusceptible children, we must realize that such a child would not only have been exposed to thimerosal mercury from being vaccinated but the child would be expected to have also prenatal and postnatal exposure to the following: methylmercury, dental amalgam mercury, and thimerosal/ ethyl-mercury . It would be reasonable to believe that the vaccination mercury could trigger an autism response.

Thus, it is of interest to review these possible exposures to mercury.

i) **Methyl mercury** from the fish that the mother consumed during pregnancy.

ii) **Methyl mercury** from the chicken that the mother consumed during pregnancy, since chicken feed often contains fish bones and fish remains.

In a 2003 examination of the National Health and Nutrition Examination Survey, a cross-sectional survey of the non-institutionalized U. S. population, it was found that approximately 8% of women had blood mercury concentrations higher than the U. S. EPA recommended reference dose of 5.8 $\mu\text{g/L}$ (CDC 2003). Thus, mercury exposure continues to be a public health concern. While the greatest mercury exposure to the general population is through mercury contained in dental amalgams (NRC, 2000), the evidence to date indicates that exposure to methyl mercury causes the most toxic responses

Methyl mercury (MeHg), a form of organic mercury, is found almost exclusively in fish. Approximately 95% to 99% of ingested methyl mercury is absorbed by the gastrointestinal tract, after which much of it combines with the amino acid cysteine and is rapidly transported to the blood-brain barrier. The cysteinyl methyl mercury structure is believed to be analogous in chemical structure to that of the amino acid methionine. Thus, methyl mercury is transported across the blood-brain barrier by the protein that transports methionine. Once methyl mercury is deposited in the brain, it is slowly demethylated to mercuric mercury (Vahter et al., 1994) that **strongly binds** to brain proteins.

iii) **Elemental mercury** from amalgams that the mother had present or had been installed during pregnancy.

Dental amalgams, so called “silver” fillings, emit elemental mercury vapor. (Skare and Engqvist, 1994). This dental restoration material can contain up to about 50% mercury. The average person with the average number of amalgam surfaces emits and retains about 10 μg

mercury per day from such amalgams. The elemental mercury vapor is rapidly moved from the mouth cavity to the lungs where it is rapidly absorbed, enters the blood and transported to the tissues and brain. Because elemental mercury is fat-soluble, it is able to pass through the blood-brain barrier. Once it gets into the brain, it is rapidly oxidized to inorganic mercury (Hg^{+2}), which binds strongly and tenaciously to brain protein and thus remains in the brain. The health effects of dental amalgams include increases in antibiotic resistance (Rowland et al. 1984; Summers et al., 1993), the in vitro destruction of snail brain neurons (Leong et al., 2001), and hypersensitivity problems in approximately 0.5 to 15% of the population. Sweden, Britain, and Germany have prohibited dental amalgams restorations for pregnant woman and young children. These two vulnerable groups have also been cautioned to limit their ingestion of seafood.

iv.) **Ethyl mercury** from the thimerosal of vaccines that the mother may have been given during her pregnancy. Thimerosal from vaccines is metabolized to ethyl mercury. While ethyl mercury is analogous in chemical structure to methyl mercury, its toxicokinetic properties are quite different. Most importantly, thimerosal is metabolized very quickly to ethyl mercury which is transported in the blood to the tissues, crosses the blood-brain barrier, and is converted to mercuric mercury in the brain at a far faster rate than methyl mercury (Burbacher et al., 2005).

Once any form of mercury enters the brain and is converted to mercuric mercury, it is not easily eliminated from the brain, despite estimates of brain mercury half-life data appearing in scientific literature. In fact, inorganic mercury, the mercuric species, formed in the brain cannot easily leave and therefore it will be trapped and accumulate in the brain (Magos et al., 1985, NRC, 2000). Mercuric mercury has been used for over 60 years as an in vitro and in vivo enzyme inhibitor, as every first year biochemistry student soon learns. It is not innocuous even when formed in vivo.

An example of methylmercury demethylation and the powerful retention of the resulting mercuric mercury in the brain is as follows: During August of 1969, a New Mexico farmer fed his hogs grain seeds treated with a fungicide containing methyl mercury. A month later, a hog was killed and the family ate it for three months. In December, several of the children became ill with neurological disorders. Deaths occurred. However, one daughter, eight years old at the time of exposure, lived until the age of 29. An autopsy determined that 100% of the mercury in her brain was inorganic. In her cerebellum, the total mercury concentration was 100 times greater than controls, even though it was about 21 years after the original exposure (Davis et al., 1994). In another study, exceedingly high levels of mercury were demonstrated in the human brain and other organs 17 years after metallic mercury exposure (Opitz et al., 1996). In summary, once mercury converts to mercuric mercury in the brain, elimination of the mercury from the brain is extremely slow if it occurs at all. It is pertinent to remember that mercuric mercury has a very high affinity for sulfhydryl groups and it is used extensively in vitro as a potent, powerful inhibitor of enzyme activity and other protein functions. Mercuric mercury, although unable to pass through the blood brain barrier, does accumulate in large amounts in the brain as the metabolic end product of the other forms of mercury that do pass through the blood brain barrier (Vahter et al., 1994). It is noteworthy that the developing brains of young children are particularly vulnerable to damage by organic (ethyl mercury and methyl mercury) and elemental mercury. In this regard, it must be emphasized; that children are not little adults. They are physically immature developing human beings with substantial metabolic differences as compared to adults. A good example of this is that at birth the blood brain-barrier is not completely developed.

One of the most common forms of mercury exposure for humans is methyl mercury by the ingestion of fish. It occurs as a result of mercury vapor and mercuric mercury becoming airborne when emitted from soil, mountains, chimneys of fossil fuel-burning utility plants and incinerators. The mercury in the air then enters rivers, bays, and oceans by means of rain and other phenomena, including leaching from the soil. Bacteria methylate the Hg^0 and Hg^{+2} in the water and in the sediment of oceans, rivers, and lakes. Unicellular organisms take up the MeHg. Then, larger organisms take up the unicellular organisms. Through a process called biomagnification, small fish containing small amounts of MeHg are eaten by larger predators, such as swordfish, tuna, shark, and tilefish which then will contain large amounts of methyl mercury. Humans then consume these larger predator fish with substantial concentrations of methyl mercury. In this regard, it is noteworthy that methyl mercury is a proven human teratogen and is particularly hazardous to pregnant women and the developing brains of young children. (The medical definition of teratogen is “monster forming”.) Two methyl mercury public health disasters have been well documented. These two public health disasters occurred in Japan and Iraq and demonstrated that methyl mercury can cross the placenta and be detrimental to the developing brains of embryos and young children.

Pink Disease- The fact that not all children who were exposed to mercurous mercury-containing teething powder came down with Pink disease and the fact that the mortality of the children was not 100% demonstrated that the genetic hypersusceptibility of some children to mercury played a significant role with respect to the nature and extent of the injury.

Between 1850 and 1950, a common syndrome, known as Pink Disease, acrodynia, erythredema, or Feer-Swift Disease (Dally, 1997), affected young children. As noted earlier, a teething powder containing mercurous salts caused this condition. Affected babies and toddlers

were unsightly and miserable. Their skin was bright pink or red in color. They were photophobic with “raw beef” hands and feet. They had peeling skin and gangrene in their extremities. The mortality was 5.5% to 33.3%. At the time, the symptoms were thought to be due to a viral disease or a nutritional deficiency. In 1920, when Pink Disease was discovered in three areas of the United States, investigators identified a mercurous mercury compound in a teething powder (as the cause. *Curiously, established, conservative medicine for years denied that Pink Disease was caused by mercury. In the mid 1950’s, however, the federal government finally banned the use of mercurous mercury in teething powder and new cases of Pink Disease disappeared.* As a final note, the fact that neither morbidity nor mortality of the children was 100% demonstrates that the genetic hypersusceptibility of some children to mercury plays a significant role with respect to the nature and extent of the injury.

Thimerosal and childhood vaccines- Thimerosal is an organic mercurial metabolized quickly to ethyl mercury (Takeda and Ukita, 1969; Magos et al., 1985; Suzuki et al., 1963). It has been used as a preservative in numerous childhood vaccines. The Thimerosal molecule contains approximately 50% mercury and 54% ethyl mercury. Some children received 187.5 mcg ethyl mercury from Thimerosal-containing vaccines during the first 14 weeks of life (Clements et al., 2001). Thimerosal is given as repeated doses equivalent to 5-20 mcg ethyl mercury over the first six months of life (Halsey, 1999). In pre-term infants, blood mercury levels after only one hepatitis B vaccination increased more than 10-fold to levels above U.S. EPA guideline of 5 mcg per liter of blood (Stajich et al., 2000). Mercury contained within the vaccines is administered as a bolus injection resulting in an exceedingly high mercury compound being injected at one time. The half time of such injected mercury in the serum of *normal* human infants was estimated to be seven days (Pichichero et al., 2002). However, recent evidence obtained from primates seems

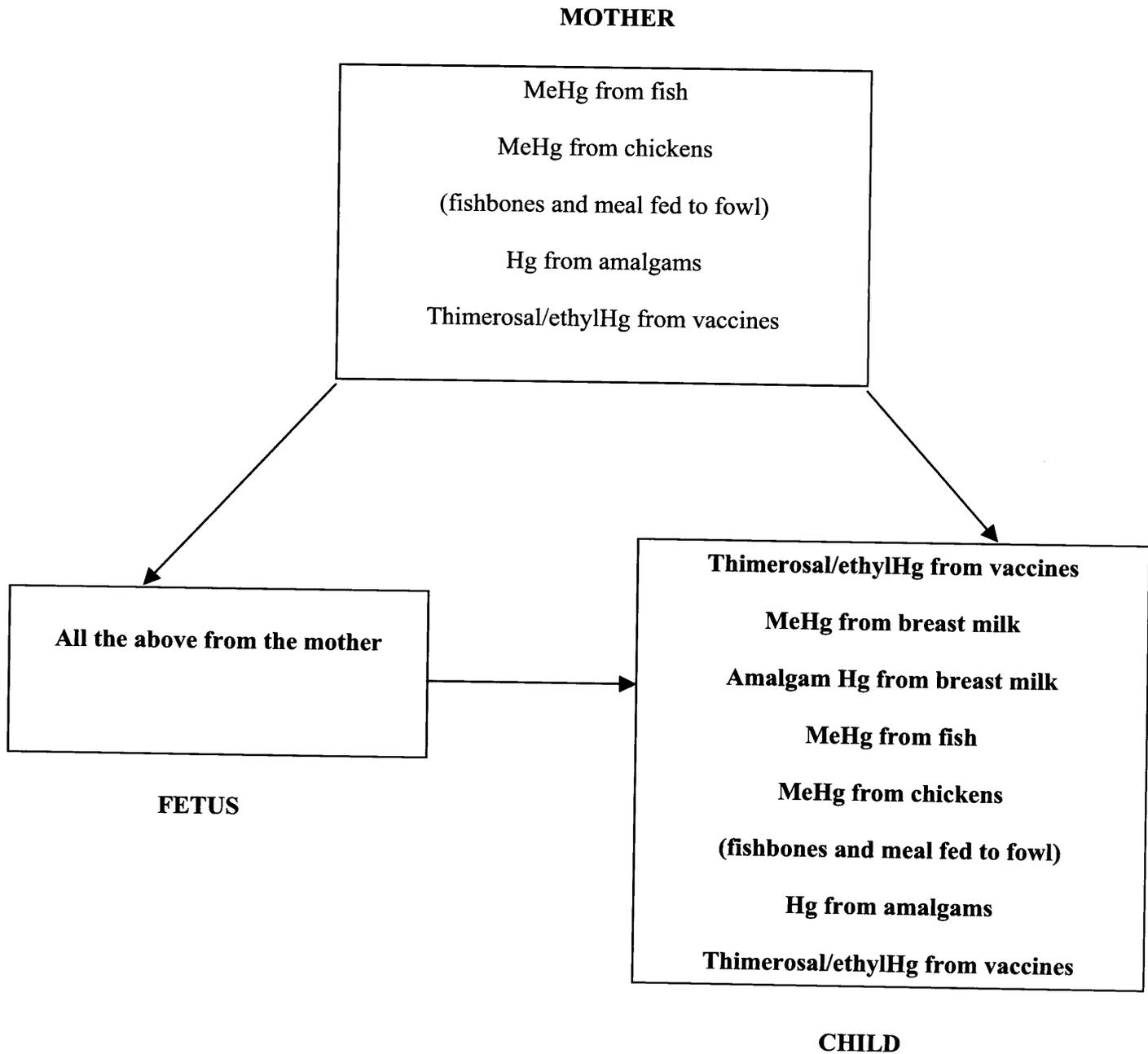
to indicate that a disproportionate amount of mercury from ethylmercury finds its way to the brain when compared to oral ingestion of methyl mercury (Burbacher et al., 2005).

Because of Burbacher's and other papers, it is often incorrectly stated that the toxicities of methylmercury and ethylmercury are very different. This statement is incorrect. Based on the data of Burbacher et al. (2005) and others, the toxicokinetics, not the toxicities, of methyl and ethyl mercury differ. There is not sufficient evidence to say that in the human the toxicities of methylmercury and ethylmercury are different.

While the chemical structure of ethyl mercury, $\text{CH}_3\text{CH}_2\text{Hg}^+$, is similar to methyl mercury, CH_3Hg^+ , its toxicokinetics are markedly different. In Burbacher et al. (2005), the authors demonstrated that when infant monkeys were given Thimerosal, the ethyl mercury in the brain was converted to inorganic mercury at a rate seven times faster than occurred for methyl mercury. Once formed in the brain, inorganic mercury, as noted above, is bound and held firmly by brain proteins because of its great affinity for thiol groups of proteins. The initial $t_{1/2}$ (half time) of ethyl mercury in the blood of the infant monkeys was 2.1 days. The terminal $t_{1/2}$ in blood was 8.6 days. For methyl mercury the $t_{1/2}$ was 21.5 days. While methyl mercury kinetics indicated a one-compartment model, the kinetics for ethyl mercury indicated a two-compartment model (Burbacher et al., 2005). For this reason, the disposition kinetics and demethylation rates differ. Because of these reasons, the toxicokinetics of methylmercury and ethylmercury differ. However, this does not mean the toxicity differs.

The maternal and other sources of mercury exposure for an infant is shown in Fig 4.

Figure 4. Influence of the mother and other sources for mercury exposure of infants



The mercury public health disasters- In Japan, a chemical factory dumped mercury wastes into a river that emptied into Minamata Bay. Mercury concentrations were found to be high in the water of the Bay and large amounts of methyl mercury were found in the fish. Unfortunately, by the time of this discovery, many people living near the Bay had suffered central nervous system disorders (Tsubaki and Irukayama, 1977). In fact, the word “minamata” is now used in Japan as a word synonymous with idiot.

Beginning in the 1960s, over 6,500 individuals were hospitalized with methyl mercury poisoning and numerous deaths occurred in the Iraqi mercury disaster (Bakir et al., 1973). These two public health disasters in Japan and Iraq demonstrate that methyl mercury can cross the placenta and be detrimental to the developing brains of embryos and young children. The disasters stimulated the U.S. government to sponsor two studies to investigate the effects of low-level exposure of methyl mercury. Specifically, the studies focused on the impact of the exposure with respect to the intelligence of children born to mothers from populations eating fish as their primary source of protein. The two studies, the Seychelles Island study (Myers et al., 2000; Myers et al., 2003) and the Faeroe Island study (Grandjean et al., 1997a, 1997b; Grandjean, 1999) had contrasting results. The Faeroe Island study showed cognitive deficits in seven-year-old children who had prenatal exposure to methyl mercury (Grandjean et al., 1997a). The Seychelles Island study, on the other hand, initially showed no harmful effects of methyl mercury (Myers et al., 2000).

A White House conference was called to evaluate the discrepancies in the two studies. (The writer of this report was invited and participated in this conference). Once some method changes were made, intelligence deficits were also found subsequently in some Seychelle Island children (Myers et al., 2003). The New Zealand study (Kjellstrom et al., 1986, 1989), has now

confirmed the Faeroe Island study. In these circumstances, it is clear that low level methyl mercury exposure can be detrimental to the developing brain of young children. See also Trasande et al. (2006), wherein the authors forcefully warn of the adverse public health and economic consequences of methyl mercury toxicity.

In another study, Hightower and Moore (2003), the authors reported a group of wealthy persons (CEOs, attorneys, and physicians) who presented to their physicians with central nervous system complaints. Their histories disclosed that they could afford and ate expensive fish, such as shark, swordfish, and tuna, almost exclusively as their protein source. These fish are predators and contain high concentrations of methyl mercury. After six months on a seafood-free diet, the patients returned to normal health.

6. A highly plausible theory is that thimerosal and its biotransformant ethylmercury cause injury to the developing brains of human embryos and young children.

Evidence for thimerosal/ethylmercury being implicated in the etiology of autism is summarized below. Each piece of evidence alone leaves some doubt but taken all together the evidence implicates thimerosal/ethylmercury in the etiology of some of the autism spectrum disorders. Another way of saying this is that in some specifically susceptible subset of infants who received mercury-containing vaccines on the US vaccination schedule in place from roughly 1991 to 2003, the ethylmercury probably caused the symptoms of autism in many of them.

First, Adams et al. (2007) demonstrated that teeth from autistic children contain more mercury than those from non-autistic children.

Second, Holmes et al (2003) demonstrated that the hair from the first haircut of autistic children contained statistically significant less mercury than that of control children. This would be expected if autistic children had a mercury efflux disorder; that is they could not move

mercury out of their cells. This would result in decreased blood and hair mercury (but not tooth mercury). This work has been criticized unjustly. Some of the reasons for this criticism has been the mercury analyses were performed by a commercial lab (this lab is certified by the FDA and probably performs more mercury analysis each year than any other lab including academic labs in the world. Another criticism was that the research was performed by individuals not having research experience and not recognized for their research expertise. Yet, one of them was and is funded by NIH grants and was the head of a university chemistry department. The hair analysis of controls was higher than the normal American population. The Holmes et al., study has been confirmed by the MIT group. (Hu et al. 2003)

Third, Bradstreet et al. (2003) demonstrated that autistic children given the chelating agent DMSA excreted more mercury in their urine than did control children.

The Nataf et al., (2006) study confirmed the Bradstreet et al results. In addition Nataf et al., found that the mercury-specific porphyrinuria increased in autistic children and that there was a marked decrease in the mercury-specific porphyrinuria of autistic children after chelation of mercury using DMSA.

Fourth, the most beneficial treatment for autism as reported by the parents of autistic children was chelation therapy using DMSA (meso-2,3-dimercaptosuccinic acid) as reported in a study by the Autism Research Institute in San Diego, CA (2006). Although this study has been criticized, such a clinical trial is being planned or has begun at the NIH.

Fifth, autoimmune disease sensitive mice exposed to mercury after birth develop enlarged brains and autistic-like symptoms. (Hornig et al., 2004).

Sixth, there is evidence for postnatal loss of brain cells in autism, particularly in the cerebellum. Courchesne (2005) at p. 584 and references cited there.

To repeat, each piece of evidence alone leaves some doubt but taken all together the evidence implicates thimerosal/ethylmercury as the likely precipitating agent in the etiology of some of the autism spectral disorders.

6- Conclusions

Thimerosal and its biotransformant ethylmercury cause injury to the developing brains of human embryos and young children by interfering with brain development during highly specific windows of brain development. These windows of susceptibility may occur during the prenatal and/or postnatal periods. These windows are vulnerable and open at very specific times.

Thus, thimerosal induces symptoms of autism spectrum disorders in some subsets of children. There is growing importance of neuroinflammatory events in the development of autism. All of this indicates the need for closer examination and better correlations of past research on the genetics and environmental factors causing autism. One of these environmental factors is surely thimerosal.

A handwritten signature in black ink that reads "H. Vashen Aposhian". The signature is written in a cursive style and is positioned to the left of a vertical line.

August 30, 2007

REFERENCES

- Adams JB, Romdalvik J, Ramanujam VM, Legator MS.
Mercury, lead, and zinc in baby teeth of children with autism versus controls.
J Toxicol Environ Health A. 2007 Jun;70(12):1046-51.
- Agrawal A, Kaushal P, Agrawal S, Gollapudi S, Gupta S
Thimerosal induces TH2 responses via influencing cytokine secretion by human dendritic cells
J. Leukoc. Biol. 2007 Feb;81(2):474-82. Epub 2006 Nov 1
- Aposhian HV, Aposhian MM.
Arsenic toxicology: five questions.
Chem Res Toxicol. 2006 Jan;19(1):1-15. Review.
- Atchison WD, Hare MF.
Mechanisms of methylmercury-induced neurotoxicity.
FASEB J. 1994 Jun;8(9):622-9. Review.
- Bakir F, Damluji SF, Amin-Zaki L, Murtadha M, Khalidi A, Al-Rawi NY, Tikriti S, Dhahir HI
Methylmercury Poisoning in Iraq: An Interuniversity Report
Science;181(96): 230-41
- Bauman ML
Microscopic Neuroanatomic Abnormalities in Autism
Pediatrics;87(5 Pt 2): 791-6
- Bradstreet J, Geier DA, Kartzinel JJ, Adams JB, Geier MR
A Case-Control Study of Mercury Burden in Children with Autistic Spectrum Disorders
Journal of American Physicians and Surgeons;8(3): 76-9
- Burbacher TM, Rodier PM, Weiss B
Methylmercury Developmental Neurotoxicity: A Comparison of Effects in Humans and Animals
Neurotoxicology and Teratology;12: 191-202

- Burbacher TM, Sackett GP, Mottet NK
Methylmercury Effects on the Social Behavior of *Macaca fascicularis* Infants
Neurotoxicology and Teratology;12: 65-71
- Burbacher TM, Shen DD, Lalovic B, Grant KS, Sheppard L, Damian D, Ellis S, Liberato N
Chronic maternal methanol inhalation in nonhuman primates (*Macaca fascicularis*): Exposure and toxicokinetics prior to and during pregnancy
NEUROTOXICOLOGY AND TERATOLOGY;26(2): 201-21
- Burbacher TM, Shen DD, Liberato N, Grant KS, Cernichiari E, Clarkson T
Comparison of blood and brain mercury levels in infant monkeys exposed to methylmercury or vaccines containing thimerosal.
Environ Health Perspect. 2005 Aug;113(8):1015-21
- CDC
Vaccination Coverage Among Children Entering School - United States, 2002-03 School Year
Morbidity and Mortality Weekly Report;52(33): 791-3
- Charleston JS, Body RL, Mottet NK, Vahter ME, Burbacher TM
Autometallographic determination of inorganic mercury distribution in the cortex of the calcarine sulcus of the monkey *Macaca fascicularis* following long-term subclinical exposure to methylmercury and mercuric chloride.
Toxicol Appl Pharmacol. 1995 Jun;132(2):325-33
- Charleston JS, Bolender RP, Mottet NK, Body RL, Vahter ME, Burbacher TM
Increases in the number of reactive glia in the visual cortex of *Macaca fascicularis* following subclinical long-term methyl mercury exposure.
Toxicol Appl Pharmacol. 1994 Dec;129(2):196-206.
- Cheslack-Postava K, Fallin MD, Avramopoulos D, Connors SL, Zimmerman AW, Eberhart CG, Newschaffer CJ
beta2-Adrenergic receptor gene variants and risk for autism in the AGRE cohort
Mol Psychiatry. 2007 Mar;12(3):283-91. Epub 2007 Jan 2
- Cheung MK, Verity MA
Experimental Methyl Mercury Neurotoxicity: Locus of Mercurial Inhibition of Brain Protein Synthesis In Vivo and In Vitro
Journal of Neurochemistry;44(6): 1799-1808

Clarkson TW
The Three Modern Faces of Mercury
Environmental Health Perspectives;110(Suppl 1): 11-23

Clarkson TW, Magos L
The toxicology of mercury and its chemical compounds.
Crit Rev Toxicol. 2006 Sep;36(8):609-62

Clarkson TW, Magos L, Myers GJ
The toxicology of mercury--current exposures and clinical manifestations
N. Engl. J. Med. 2003 Oct 30;349(18):1731-7

Clements CJ, Ball LK, Ball R, Pratt RD
Thiomersal in Vaccines: Is Removal Warranted?
Drug Safety;24(8): 698-99

Committee on the Toxicological Effects of Methylmercury
Toxicological Effects of Methylmercury
National Research Council;

Connors SL, Crowell DE, Eberhart CG, Copeland J, Newschaffer CJ, Spence SJ, Zimmerman
AW
beta2-adrenergic receptor activation and genetic polymorphisms in autism: data from dizygotic
twins
J Child Neurol. 2005 Nov;20(11):876-84

Costa LG, Aschner M, Vitalone A, Syversen T, Soldin OP
Developmental neuropathology of environmental agents.
Annu Rev Pharmacol Toxicol. 2004;44:87-110.

Counter S, Buchanan L, Ortega F
Neurocognitive screening of mercury-exposed children of Andean gold miners.
Int J Occup Environ Health. 2006 Jul-Sep;12(3):209-14.

Courchesne E, Redcay E, Morgan JT, Kennedy DP.
Autism at the beginning: microstructural and growth abnormalities underlying the cognitive and
behavioral phenotype of autism.

Dev Psychopathol. 2005 Summer;17(3):577-97. Review.

Dally A
The Rise and Fall of Pink Disease
Society for the Social History of Medicine;10(2): 291-304

Davis LE, Kornfeld M, Mooney H, Fielder KJ, Haaland KY, Orrison WW, Cernichiari E, Clarkson TW
Methylmercury Poisoning: Long-Term Clinical, Radiological, Toxicological, and Pathological Studies of an Affected Family
Annals of Neurology;35(6): 680-6

Gilbert SG, Rice DC, Burbacher TM
Fixed interval/fixed ratio performance in adult monkeys exposed in utero to methylmercury.
Neurotoxicol Teratol. 1996 Sep-Oct;18(5):539-46.

Grandjean P
Mercury Risks: Controversy or Just Uncertainty
Viewpoint;114:512-5

Grandjean P
Mercurial Uncertainties in Environmental Health
Annual of the New York Academy of Science;837:239-45

Grandjean P, Budtz-Jorgensen E, White R, Jorgensen PJ, Weihe P, Debes F, Keiding N
Methylmercury Exposure Biomarkers as Indicators of Neurotoxicity in Children Aged 7 Years
American Journal of Epidemiology;150(3):301-5

Grandjean P, Weihe P, White RF, Debes F
Cognitive Performance of Children Prenatally Exposed to "Safe" Levels of Methylmercury"
Environmental Research;77:165-72

Grandjean P, Weihe P, White RF, Debes F, Araki S, Yokoyama K, Murata K, Sorensen N, Dahl R, Jorgensen PJ
Cognitive deficit in 7-year-old children with prenatal exposure to methylmercury
Neurotoxicology and Teratology;19(6):417-28

Halsey NA
Limiting Infant Exposure to Thimerosal on Vaccines and Other Sources of Mercury
Journal of American Medical Association;282:1763-1766

Heyer NJ, Bittner AC Jr, Echeverria D, Woods JS
A cascade analysis of the interaction of mercury and coproporphyrinogen oxidase (CPOX)
polymorphism on the heme biosynthetic pathway and porphyrin production.

Toxicol Lett. 2006 Feb 20;161(2):159-66. Epub 2005 Oct 7

Hightower JM, Moore D
Mercury Levels in High-End Consumers of Fish
ENVIRONMENTAL HEALTH PERSPECTIVES;111(4): 604-8

Holmes AS, Blaxill MF, Haley BE
Reduced Levels of Mercury in First Baby Haircuts of Autistic Children
International Journal of Toxicology;22(4): 277-85

Hornig, M, et al
Neurotoxic Effects of Postnatal Thimerosal Are Mouse Strain Dependant
Molecular Psychiatry 2004:1-13

Hu, L et al.
Neutron activation analysis of hair samples for the identification of autism.
Poster presentation:Trans Am Nucl Soc 2003;89

Institute of Medicine
Immunization Safety Review Committee: Thimerosal-Containing Vaccines and
Neurodevelopmental Outcomes Public Meeting
National Academy of Sciences;

Kern JK, Jones AM
Evidence of toxicity, oxidative stress, and neuronal insult in autism.
J Toxicol Environ Health B Crit Rev. 2006 Nov-Dec;9(6):485-99

Kjellstrom T, Kennedy P, and Mantell C
Physical and mental development of children with prenatal exposure to mercury from fish. Stage
1: Preliminary tests at age 4.
Report 3080. Solna, Sweden: National Swedish Environmental Protection Board.

Kjellstrom T, Kennedy P, Wallis S, Stewart A, Friberg L, and Lind B.
Physical and mental development of children with prenatal exposure to mercury from fish. Stage II: Interviews and psychological tests at age 6.
Report 3642. Solna, Sweden: National Swedish Environmental Protection Board.

Leong CW, Syed NI, Lorscheider FL
Retrograde degeneration of neurite membrane structural integrity of nerve growth cones following in vitro exposure to mercury
Neuroreport;12:733-8

Magos L, Brown AW, Sparrow S, Bailey E, Snowden RT, Skipp WR
The comparative toxicology of ethyl- and methylmercury
Archives of Toxicology;57:260-67

Miller MT, Stromland K.
Teratogen update: thalidomide: a review, with a focus on ocular findings and new potential uses. Teratology. 1999 Nov;60(5):306-21. Review.

Morton NE.
Genetic epidemiology of hearing impairment. Ann N Y Acad Sci. 1991;630:16-31. Review. No abstract available.

Mutter J, Naumann J, Guethlin C.
Comments on the article "the toxicology of mercury and its chemical compounds" by Clarkson and Magos (2006)."
Crit Rev Toxicol. 2007 Jul;37(6):537-49.

Myers GJ, Davidson PW, Cox C, Shamlaye CF, Palumbo D, Cernichiari E, Sloane-Reeves J, Wilding GE, Kost J, Huang L, Clarkson TW
Prenatal methylmercury exposure from ocean fish consumption in the Seychelles child development study
Lancet;361(9370): 1686-92

Myers GJ, Davidson PW, Palumbo D, Shamlaye C, Cox C, Cernichiari E, Clarkson TW.
Secondary analysis from the Seychelles Child Development Study: the child behavior checklist.
Environ Res. 2000 Sep;84(1):12-9.

Nataf R, Skorupka C, Amet L, Lam A, Springbett A, Lathe R
Porphyrinuria in childhood autistic disorder: implications for environmental toxicity
Toxicol Appl Pharmacol. 2006 Jul 15;214(2):99-108. Epub 2006 Jun 16

Nordberg, GF, Fowler BA, Nordberg M, Friberg LT.
Handbook on the toxicology of metals,
3d edition, Academic Press 2007

Opitz H, Schweinsberg F, Grossman T, Wendt-Gallitelli MF, Meyermann R
Demonstration of mercury in the human brain and other organs 17 years after metallic mercury
exposure
CLINICAL NEURPATHOLOGY;15(3): 139-44

Palmer RF, Blanchard S, Stein Z, Mandell D, Miller C
Environmental mercury release, special education rates, and autism disorder: an ecological study
of Texas.
Health Place. 2006 Jun;12(2):203-9

Pardo CA, Vargas DL, Zimmerman AW
Immunity, neuroglia and neuroinflammation in autism
Int Rev Psychiatry. 2005 Dec;17(6):485-95

Pichichero ME, Cernichiari E, Lopreiato J, Treanor J
Mercury concentrations and metabolism in infants receiving vaccines containing thiomersal: a
descriptive study
Lancet;360(9347): 1737-40

Rhodes MC, Seidler FJ, Abdel-Rahman A, Tate CA, Nyska A, Rincavage HL, Slotkin TA.
Terbutaline is a developmental neurotoxicant: effects on neuroproteins and morphology in
cerebellum, hippocampus, and somatosensory cortex.
J Pharmacol Exp Ther. 2004 Feb;308(2):529-37. Epub 2003 Nov 10.

Rhodes MC, Seidler FJ, Qiao D, Tate CA, Cousins MM, Slotkin TA.
Does pharmacotherapy for preterm labor sensitize the developing brain to environmental
neurotoxicants? Cellular and synaptic effects of sequential exposure to terbutaline and
chlorpyrifos in neonatal rats.
Toxicol Appl Pharmacol. 2004 Mar 1;195(2):203-17.

Rimland B
Treatment Options for Mercury/Metal Toxicity in Autism and Related Developmental
Disabilities: Consensus Position Paper
Autism Research Institute

Rowland IR, Robinson RD, Doherty, RA
Effects of Diet on Mercury Metabolism and Excretion in Mice Given Methylmercury: Role of
Gut Flora
Archives of Environmental Health;39(6): 401-8

Schober SE, Sinks TH, Jones RL, Bolger PM, McDowell M, Osterloh J, Garrett ES, Canady RA,
Dillon CF, Sun Y, Joseph CB, Mahaffey KR
Blood Mercury Levels in US Children and Women of Childbearing Age, 1999-2000
Journal of American Medical Association;289(13): 1667-74

Skare I, Engqvist A
Human exposure to mercury and silver released from dental amalgam restorations
Arch. Environ. Health. 1994 Sep-Oct;49(5):384-94

Stajich GV, Lopez GP, Harry SW, Sexson WR
Iatrogenic exposure to mercury after hepatitis B vaccination in preterm infants
Journal of Pediatrics;136: 679-81

Streit WJ, Mrak RE, Griffin WS
Microglia and neuroinflammation: a pathological perspective
J Neuroinflammation. 2004 Jul 30;1(1):14

Summers AO, Wireman J, Vimy MJ, Lorscheider FL, Marshall B, Levy SB, Bennett S, Billard L
Mercury released from dental silver" fillings provokes an increase in mercury- and antibiotic-
resistant bacteria in oral and intestinal floras of primates"
Antimicrob. Agents Chemother. 1993 Apr;37(4):825-34

Suzuki T, Miyama T, Katsunuma H
Comparative Study of Bodily Distribution of Mercury in Mice After Subcutaneous
Administration of Methyl, Ethyl and n-Propyl Mercury Acetates
Japanese Journal of Experimental Medicine;33(5): 277-82

Takeda Y, Ukita T
Metabolism of Ethylmercuric Chloride-203 Hg in Rats
Toxicology and Applied Pharmacology;17(1): 181-88

Trasande L, Landrigan PJ, Schechter C
Public health and economic consequences of methyl mercury toxicity to the developing brain
Environ. Health Perspect. 2005 May;113(5):590-6

Trasande L, Schechter CB, Haynes KA, Landrigan PJ
Mental retardation and prenatal methylmercury toxicity
Am. J. Ind. Med. 2006 Mar;49(3):153-8

Tsubaki T and Irukayama K (Eds.)
Minamata Disease: Methylmercury Poisoning in Minamata and Niigata, Japan.
Elsevier, New York. 1977

Ukita T, Takeda Y.
[Metabolic fate of mercury compounds]
Shinkei Kenkyu No Shimpo. 1969 Apr;13(1):123-9. Japanese.

Vahter M, Mottet NK, Friberg L, Lind B, Shen DD, Burbacher T
Speciation of mercury in the primate blood and brain following long-term exposure to methyl mercury
Toxicol Appl Pharmacol. 1994 Feb;124(2):221-9

Vahter ME, Mottet NK, Friberg LT, Lind SB, Charleston JS, Burbacher TM
Demethylation of methyl mercury in different brain sites of *Macaca fascicularis* monkeys during long-term subclinical methyl mercury exposure
Toxicol Appl Pharmacol. 1995 Oct;134(2):273-84

Vineis P, Kriebel D.
Causal models in epidemiology: past inheritance and genetic future.
Environ Health. 2006 Jul 21;5:21. Review.

Woods JS, Echeverria D, Heyer NJ, Simmonds PL, Wilkerson J, Farin FM
The association between genetic polymorphisms of coproporphyrinogen oxidase and an atypical porphyrinogenic response to mercury exposure in humans.
Toxicol Appl Pharmacol. 2005 Aug 7;206(2):113-20

Zerrate MC, Pletnikov M, Connors SL, Vargas DL, Seidler FJ, Zimmerman AW, Slotkin TA, Pardo CA.

Neuroinflammation and behavioral abnormalities after neonatal terbutaline treatment in rats: implications for autism.

J Pharmacol Exp Ther. 2007 Jul;322(1):16-22. Epub 2007 Mar 30.

November, 2006

CURRICULUM VITAE

H. Vasken Aposhian, PhD
Professor of Molecular and Cellular Biology
Professor of Pharmacology
University of Arizona

Place of Birth: Providence, RI

Education

B.S. Brown University, 1948 (Chemistry)

M.S. University of Rochester, 1950 (Physiological Chemistry)

Ph.D. University of Rochester, 1953 (Physiological Chemistry)

Advanced Training:

Department of Biochemistry, Stanford University School of Medicine (with Nobel Laureate Dr. Arthur Kornberg), 1959-1962

Department of Biology, Massachusetts Institute of Technology (with Dr. Paul Schimmel) -- six-month sabbatical, January 1-June 30, 1983

Department of Biology, University of California, San Diego -- six-month sabbatical as Visiting Scholar, June 1-December 30, 1990

Positions Held

- 1954-56 Instructor, Department of Pharmacology, Vanderbilt University School of Medicine
- 1956-59 Assistant Professor, Department of Pharmacology, Vanderbilt University School of Medicine
- 1959-62 USPHS Senior Research Fellow, Department of Biochemistry, Stanford University School of Medicine
- 1962-67 Associate Professor, Department of Microbiology, Tufts University School of Medicine
- 1966-75 Professor, Department of Cell Biology and Pharmacology, University of Maryland School of Medicine. (As Head of Department, 1966-72)
- 1975-present **Professor, Department of Pharmacology, College of Medicine, University of Arizona**
- 1975-83 Professor, Department of Cellular and Developmental Biology, College of Liberal Arts, University of Arizona. (Head of Department, 1975-79)
- 1983 Visiting Professor, Department of Biology, Massachusetts Institute of Technology (January 1, 1983-June 30, 1983)
- 1983-present **Professor, Department of Molecular and Cellular Biology, Faculty of Science, University of Arizona**

Research Interests

1. Metal toxicity and mechanisms of intoxication of arsenic, mercury, lead, and manganese, polymorphisms involved.
2. Biological chelation: Use of DMPS as a challenge test for mercury, arsenic and lead.
3. Arsenic detoxification and intoxication: molecular mechanisms.
4. DNA and gene delivery systems for mammalian cells and intact animals.
5. Pseudovirions.
6. Autism

Professional Societies

Society for Toxicology
American College of Toxicology
American Society of Biological Chemistry and Molecular Biology
American Society of Microbiology
American Society for the Advancement of Science, Fellow
New York Academy of Sciences
American Society for Pharmacology and Experimental Therapeutics (resigned 1976)
American Academy of Microbiology
American Association of University Professors

Awards or Honors

1959-64	USPHS Senior Research Fellowship (resigned 1962)
1959	Jane Coffin Child Fellowship (declined)
1972	Sigma Xi Annual Award for Scientific Achievement, Maryland Chapter
1974	Student Council Award for Excellence in Teaching, University of Maryland School of Medicine
1977	Invited Guest, Soviet Academy of Science, June, 1977
1977	Invited Lecturer, Al-Hazen Research Institute, Baghdad, Iraq, May, 1977
1981	Invited Speaker, Korean Biochemical Society
1985	National Academy of Science (U.S.) - Soviet Union Academy of Science Exchange Fellow for September, 1985, in Soviet Union
1985	Official Guest of Peoples' Republic of China, Academy of Science, Lecture Tour, October, 1985
1996	Official Guest of Peoples' Republic of China, Academy of Science, Evaluation of arsenic problem in Guizhou Province, November, 1996
1998	Official Guest of Peoples' Republic of China, Academy of Science, Evaluation of arsenic problem Autonomous Region of Inner Mongolia, November, 1996
1999	Official Guest of Peoples' Republic of China, Academy of Science, Treatment of Arsenic problem in Guizhou Province.

National Service (only a few are listed)

- 1970-78 Member of ad hoc committees for cancer programs and cancer construction programs of the National Cancer Institute.
- 1971 Consultant to National Cancer Institute Planning Session, Airlie Conference Center.
- 1972 Advisor on gene technology to U.S. Senator J.V. Tunney.
U.S. Environmental Protection Agency, Mercury Advisory Committee.
- 1971-73 American Cancer Society, Maryland Division - Member, Board of Directors.
- 1971-72 American Cancer Society, Maryland Division -Chairman Grants Committee.
- 1989- Consultant For Various Multinational Pharmaceutical, Consumer-Goods And Life Science Companies – Confidential.
- 1990 Lecturer - Continuing Education Committee, Society of Toxicology.
- 1990-94 Member of various ad hoc study sections, National Institutes of Health especially for the National Institute for Environmental Health Sciences.
- 1993 Councilor - Metal Section, Society of Toxicology.
- 1993 Super Fund - Agenda Workshop for Biodiversity Toxicology of Children, for the National Institute for Environmental Health Sciences.
- 1995 WAARF Arsenic Research Priority Planning Meeting,-- Mechanisms Section Chairman.
- 1995 WAARF Arsenic Grant Application Study Section.
- 1997-98 National Research Council, Committee on Toxicology, Subcommittee on Arsenic in Drinking Water, member.
- 1997 EPA Working Committee on Arsenic Carcinogenesis.
- 1998 NIEHS Methylmercury evaluation group.
- 1998 Mercury toxicity. Presentation to Committee On Government Reform, House of Representatives, Congress of the United States
- 1998 Cure Autism Now, Research Grant Committee
- 1999-2000 National Research Council, Committee on Toxicology, Committee on Mercury Toxicity, member.
- 2004 Presentation to Vaccine Committee of IOM entitled: *A Toxicologist's View of Autism and Thimerosal*
- 2005-2006 EPA, Arsenic Study Committee

International Service

- Present Research and Scientific Evaluations of Arsenic, Mercury and Other Toxic Chemicals for National Governments.
- 1992 Metal Toxicology Workshop for Physicians, Taipei Veterans Hospital Center, Taiwan.
- 1993 Superfund Workshop, Campagne de Madonna, Italy.
- 1993 Mercury Levels in Mexican Dental and Tampico Factory workers.

1994 German Government Metal Toxicology Workshop.
 1994 Arsenic Toxicity in Chile.
 1994 Hg Toxicity in Denmark.
 1996 Arsenic toxicity in China.
 1998 Arsenic toxicity in Inner Mongolia.
 1999 Arsenic toxicity in China.
 1999 Arsenic toxicity in Romania
 2000 Lead, cadmium and arsenic toxicity in children in Torreon, Mexico
 2003 WHO workshop on Child Health in Southeast Asia (Bangladesh)
 2003 US-Japan meeting on arsenic (by invitation only)
 2004 Arsenic toxicity in rural elevated Argentina
 2006 International Conference of Chelating Agents, Advisory Board

University of Arizona Service

1976-79 University Advisory Committee on Promotion and Tenure.
 1975-79 Biomedical Support Research Grants Committee.
 1975-77 Executive Committee, Cancer Center.
 1976-77;
 1979 Search Committee, Biochemistry Head.
 1977-85 University Committee to review DNA recombinant research.
 1978 Toxicology Program - Member of Executive Committee.
 1979-85 Graduate Council - Chairman, Student Affairs Subcommittee; Petitions Subcommittee.
 1987 Toxicology Faculty Search Committee, College of Pharmacy.
 1988 Molecular and Cellular Biology Faculty Search Committee, College of Arts and Sciences.
 1991-95 Chairman, Biomedical Group for Superfund Center.
 1992 Member of committee to review College of Medicine Molecular and Cellular Biology course.
 1993 Chairman of committee for five-year review of Department Head.
 1994 Environmental Quality Committee.
 1995 Medical School Neurosciences Review Committee.
 1995 Chairman, Departmental Promotion and Tenure Committee.
 1998 College of Science, Promotion and Tenure Committee.

State of Arizona Service

1989-92 Commissioner, Structural Pest Control Commission, State of Arizona.

Present Grant Support

NIEHS Superfund Project, \$181,000 per year. *In vivo* and *in vitro* metabolism of arsenic.
 Autism Research Institute, \$40,000 per year. Autism Biomarkers.

Other

Paid consultant at various times for various multinational pharmaceutical or consumer product companies.

Bibliography

(Does not include abstracts of papers presented at international or national meetings or seminars and lectures given at various institutions.)

Chowdhury UK, Zakharyan RA, Hernandez A, Avram MD, Kopplin MJ, and Aposhian HV. Glutathione-S-transferase-omega [MMA(V) reductase] knockout mice: Enzyme and arsenic species concentrations in tissues after arsenate administration. *Clin Res Toxicol* 216:446-447 (2006).

Aposhian HV and Aposhian MM Arsenic Toxicology: Five Questions. *Chem Res Toxicol* 19:1-15 (2006).

Zakharyan RA, Tsaprailis G, Chowdhury UK, Hernandez A, and Aposhian HV. Interactions of Sodium Selenite, Glutathione, Arsenic Species and Omega Class Human Glutathione Transferase. *Chem Res Toxicol* 18:1287-1295 (2005).

Aposhian HV, Zakharyan RA, Avram MD, Sampayo-Reyes A, and Wollenberg M L. A review of the enzymology of arsenic metabolism and a new potential role of hydrogen peroxide in the detoxication of the trivalent arsenic species. *Toxicol and Applied Pharmacol* 198:327-335 (2004).

Marnell LL, Garcia-Vargas GG, Chowdhury UK, Zakharyan RA, Walsh B, Avram, MD, Kopplin MJ, Cebrian ME, Silbergeld EK, and Aposhian HV. Polymorphisms in the human monomethylarsonic acid (MMA V) reductase/hGSTO1 gene and changes in urinary arsenic profiles. *Chem Res Toxicol* 16:1507-1513 (2003).

Aposhian HV, Zakharyan RA, Healy SM, Wildfang E, Petrick JS, Sampayo-Reyes A, Boar PG, Carter DE, Guha Mazumder DN, and Aposhian MM. Enzymology and toxicity of inorganic arsenic. In *Arsenic Exposure and Health Effects V: Proceedings of the Fifth International Conference on Arsenic Exposure and Health Effects* (2002), pgs. 225-240, Editors: Chappell, Abernathy, Calderon, Thomas, Elsevier Science Ltd. Oxford (2003).

Carter DE, Aposhian HV, and Gandolfi AJ. The metabolism of inorganic arsenic oxides, gallium arsenide, and arsine: a toxicochemical review. *Toxicol and Applied Pharmacol* 193:309-334 (2003).

Aposhian HV, Zakharyan RA, Avram MD, Kopplin MJ, and Wollenberg ML. Oxidation and detoxification of trivalent arsenic species. *Toxicol and Applied Pharmacol* 193:1-8 (2003).

Aposhian HV, Morgan DL, Queen HLS, Maiorino RM, and Aposhian MM. Vitamin C, Glutathione nor Lipoic Acid Did Not Decrease Brain or Kidney Mercury in Rats Exposed to Mercury Vapor. *Clinical Toxicol* 41:339-347 (2003).

Loffredo CA, Aposhian HV, Cebrian ME, Yamauchi H, and Silbergeld EK. Variability in human metabolism of arsenic. *Environ Res* 92:85-91 (2003).

Bhumasamudram J, Aposhian HV, Mash EA. An efficient synthesis of sodium dimethylarsinate-¹⁴C. *J LabelCompd Radiopharm* 46:373-377 (2003).

Aposhian HV. Elemental, mercuric and organic mercury: biological interactions and dilemmas. *PowerPlant Chem* 4:557-561 (2002).

- Radabaugh TR, Sampayo-Reyes A, Zakharyan RA, and Aposhian HV. Arsenate reductase II. Purine nucleoside phosphorylase in the presence of dihydrolipoic acid is a route for reduction of arsenate to arsenite in mammalian systems. *Chem Res Toxicol* 15:692-680 (2002).
- Liu J, Zheng B, Aposhian HV, Zhou Y, Chen M-L, Waalkes MP. Chronic arsenic poisoning from burning high arsenic containing coal in Guizhou, China. *Environ Health Perspect* 110:119-122 (2002).
- Gong Z, Jiang G, Cullen WR, Aposhian HV, and Le XC. Determination of arsenic metabolic complex excreted in human urine after administration of sodium 2,3-dimercapto-1-propane sulfonate. *Chem Res Toxicol* 15:1318-1323 (2002).
- Wildfang E, Radabaugh TR, and Aposhian HV. Enzymatic methylation of arsenic compounds. IX. Liver arsenite methyltransferase and arsenate reductase activities in primates. *Toxicol* 168:213-221 (2001).
- Zakharyan RA, Sampayo-Reyes A, Healy SM, Tsapraillis G, Board PG, Liebler DC, and Aposhian, HV. Human monomethylarsonic acid (MMAV) reductase is a member of the glutathione-S-transferase superfamily *Chem Res Toxicol* 14:1051-1057 (2001).
- Petrick JS, Jagadish B, Mash EA, and Aposhian HV. Monomethylarsonous Acid (MMAIII) and Arsenite: LD50 in Hamsters and In Vitro Inhibition of Pyruvate Dehydrogenase. *Chem Res Toxicol* 14:651-656 (2001).
- Aposhian HV and Aposhian MM. Arsenic mobilization by DMPS. In *Arsenic Exposure and Health Effects: Proceedings of the Fourth International Conference (2000) on Arsenic Exposure*. pg 397-406. Editors: Chappell, Abernathy, and Calderon. Elsevier Science Ltd.Oxford (2001).
- Aposhian HV, Gurzau ES, Le XC, Gurzau A, Zakharyan RA, Cullen WR, Healy SM, Gonzalez-Ramirez D, Morgan DL, Sampayo-Reyes A, Wildfang E, Radabaugh TR, Petrick JS, Mash EA, Aavula RB, and Aposhian MM. The discovery, importance and significance of monomethylarsonous acid (MMAIII) in urine of humans exposed to inorganic arsenic. In *Arsenic Exposure and Health Effects: Proceedings of the Fourth International Conference (2000) on Arsenic Exposure*. Editors: Chappell, Abernathy, and Calderon. Elsevier Science Ltd.Oxford (2001).
- Le XC, Lu X, Ma M, Cullen WR, Aposhian HV, and Zheng B. Speciation of key arsenic metabolic intermediates in human urine. *Anal Chem* 72:5172-5177 (2000).
- Sampayo-Reyes A, Zakharyan RA, Healy SM, and Aposhian HV. Monomethylarsonic acid reductase and monomethylarsonous acid in hamster tissue. *Chem Res Toxicol* 11:1181-1186 (2000).
- Petrick JS, Ayala-Fierro F, Cullen WR, Carter DE, and Aposhian HV. Monomethylarsonous Acid (MMAIII) is More Toxic than Arsenite in Chang Human Hepatocytes. *Toxicol Appl Pharmacol* 163:203-207 (2000).
- Aposhian HV, Gurzau ES, Le XC, Gurzau A, Healy SM, Lu X, Ma M, Yip L, Zakharyan RA, Maiorino RM, Dart RC, Tircus MG, Gonzalez-Ramirez D, Morgan DL, Avram D, and Aposhian MM. Occurrence of Monomethylarsonous acid (MMA^{III}) in Urine of Humans Exposed to Inorganic Arsenic. *Chem Res Toxicol* 13:693-697 (2000).
- Le XC, Ma M, Lu X, Cullen WR, Aposhian HV, and Zheng B. Determination of monomethylarsonous acid, a key arsenic methylation intermediate, in human urine. *Environ Health Perspect* 108:1015-1018 (2000).

Gailer JG, George GN, Pickering IJ, Prince RC, Ringwald SC, Pemberton JE, Glass RS, Younis HS, DeYoung DW, and Aposhian HV. A metabolic link between arsenite and selenite: The seleno-bis(S-glutathionyl) arsinium ion. *J Am Chem Soc* 122:4637-4639 (2000).

Aposhian HV, Zheng B, Aposhian MM, Le XC, Cebrian ME, Cullen W, Zakharyan RA, Ma M, Dart RC, Cheng Z, Andrewes P, Yip L, O'Malley GF, Maiorino RM, Van Voorhies W, Healy SM, and Titcomb A. DMPS-arsenic challenge test: II. Modulation of arsenic species, including monomethylarsonous acid (MMA^{III}), excreted in human urine. *Toxicol Appl Pharmacol* 164:74-83 (2000).

Radabaugh TR and Aposhian H. Enzymatic Reduction of Arsenic Compounds in Mammalian Systems: Reduction of Arsenate to Arsenite by Human Liver Arsenate Reductase, *Chem Res Toxicol* 13:26-30 (2000)

Zakharyan RA and Aposhian H V. Enzymatic Reduction of Arsenic Compounds in Mammalian Systems: The Rate Limiting Enzyme of Rabbit Liver Arsenic Biotransformation is MMA^V Reductase. *Chem Res Toxicol* 12:1278-1283 (1999).

Aposhian HV, Zakharyan RA, Wildfang EK, Healy SM, Gailer J, Radabaugh TR, Bogdan GM, Powell LA, and Aposhian MM. How is inorganic arsenic detoxified? In *Arsenic Exposure and Health Effects: Proceedings of the Third International Conference on Arsenic Exposure and Health Effects, July 12-15, 1998, San Diego, California*, pgs. 289-297, Editors: Chappell WR, Abernathy CO, and Calderon RL. Elsevier Science Ltd. Oxford (1999).

Bogdan GM, Goud GN, Sampayo-Reyes A, Porath J, Aposhian MM, and Aposhian HV. Arsenic binding proteins of mammalian systems: II. Purification of a 450 kDa arsenite-binding protein from rabbit liver. Submitted.

Healy SM, Wildfang E, Zakharyan RA, and Aposhian HV. Diversity of inorganic arsenite biotransformation. *Biol. Trace Elem Res* 68:249-266 (1999).

Zakharyan RA and Aposhian HV. Arsenite methylation by methylvitamin B₁₂ and glutathione does not require an enzyme. *Toxicol Appl Pharmacol* 154:287-291 (1999).

Zakharyan RA, Ayala-Fierro F, Cullen WR, Carter DE, and Aposhian HV. Enzymatic methylation of arsenic compounds: VII - MMA^{III} is the substrate for MMA methyltransferase of rabbit liver and human hepatocytes. *Toxicol Appl Pharmacol* 158:9-15, (1999).

Echeverria D, Aposhian HV, Woods JS, et al. Neurobehavioral effects from exposure to dental amalgam Hg degrees: New distinctions between recent exposure and Hg body burden. *FASEB J* 12:971-980 (1998).

Gonzalez-Ramirez D, Zuniga-Charles M, Narro-Juarez A, Molina-Recio Y, Hurlbut KM, Dart RC, and Aposhian HV. DMPS (2,3-dimercaptopropane-1-sulfonate, Dimaval) decreases the body burden of mercury in humans exposed to mercurous chloride. *J Pharmacol Exp Ther* 287:8-12 (1998).

Healy SM, Casarez EA, Ayala-Fierro F, and Aposhian HV. Enzymatic methylation of arsenic compounds: V. Arsenite methyltransferase activity in tissues of mice. *Toxicol Appl Pharmacol* 148:65-70 (1998).

Wildfang E, Zakharyan RA, and Aposhian HV. Enzymatic methylation of arsenic compounds: VI. Arsenite and methylarsonic acid methyltransferase kinetics. *Toxicol Appl Pharmacol* 152:366-375 (1998).

Aposhian HV. Enzymatic methylation of arsenic species and other new approaches to arsenic toxicity. *Annu Rev Pharmacol Toxicol* 37:397-419 (1997).

Aposhian HV, Arroyo A, Cebrian ME, Del Razo LM, Hurlbut KM, Dart RC, Gonzalez-Ramirez D, Kreppel H, Speisky H, Smith A, Gonsebatt ME, Ostrosky-Wegman P, and Aposhian MM. DMPS-arsenic challenge test: I. Increased urinary excretion of monomethylarsonic acid in humans given dimercaptopropane sulfonate. *J Pharmacol Exp Ther* 277:938-944 (1997).

Aposhian HV. Mobilization of mercury and arsenic in humans by sodium 2,3-dimercapto-1 propane sulfonate (DMPS). *Environ Health Perspect* 106:1017-1025 (1998).

Cianciola ME, Echeverria D, Martin MD, Aposhian HV, and Woods JS. Epidemiologic assessment of measures used to indicate low-level exposure to mercury vapor (Hg⁰). *J Toxicol Environ Health* 52:19-33 (1997).

Aposhian HV. Enzymatic methylation of arsenic species and other new approaches to arsenic toxicity. *Annu Rev Pharmacol Toxicol* 37:397-419 (1997).

Aposhian HV, Zakharyan RA, Wu Y, Healy S, and Aposhian MM. Enzymatic methylation of arsenic compounds: II. An overview. In Abernethy C, Calderon R, and Chappell W, eds., *Human Health II*, London: Chapman & Hall (1997).

Healy S, Zakharyan RA, and Aposhian HV. Enzymatic methylation of arsenic compounds: IV. In vitro and in vivo deficiency of the methylation of arsenite and monomethylarsonic acid in the guinea pig. *Mutation Res* 386:229-239 (1997).

Aposhian HV, Arroyo A, Cebrian ME, Del Razo LM, Hurlbut KM, Dart RC, Gonzalez-Ramirez D, Kreppel H, Speisky H, Smith A, Gonsebatt ME, Ostrosky-Wegman P, and Aposhian MM. DMPS-Arsenic challenge test: I. Increased Urinary Excretion of Monomethylarsonic acid in humans given dimercaptopropane sulfonate. *J Pharmacol Exp Ther* 282:192-200 (1997).

Aposhian MM, Maiorino RM, Xu Z, and Aposhian HV. Sodium 2,3-dimercapto-1-propanesulfonate (DMPS) treatment does not redistribute lead or mercury to the brain of rats. *Toxicol* 109:49-55 (1996).

Zakharyan RA, Wildfang E, and Aposhian HV. Enzymatic methylation of arsenic compounds: III. The marmoset and tamarin, but not the rhesus, monkeys are deficient in methyltransferases that methylate inorganic arsenic. *Toxicol Appl Pharmacol* 140:77-84 (1996).

Aposhian HV. Arsenic toxicology: Does methylation of arsenic species have an evolutionary significance? In Collery, P., Corbella, J., Domingo, J. L., and Etienne, J.-C., Llobet, J. M., eds. *Metal Ions in Biology and Medicine*, vol. 4, Proceedings of the Fourth International Symposium on Metal Ions in Biology and Medicine held in Barcelona (Catalonia) Spain, May 19-22, pp. 399-401, (1996).

Maiorino RM, Gonzalez-Ramirez D, Zuniga-Charles M, Xu Z, Hurlbut KM, Aposhian MM, Dart RC, Woods JS, Ostrosky-Wegman P, Gonsebatt ME, and Aposhian HV. Sodium 2,3-dimercaptopropane-1-sulfonate challenge test for mercury in humans. III. Urinary mercury after exposure to mercurous chloride. *J Pharm Exp Ther* 227:938-944 (1996).

Aposhian MM, Maiorino RM, Xu Z, and Aposhian HV. Sodium 2,3-dimercapto-1-propanesulfonate treatment does not redistribute mercury or lead to the brain of rats. In Collery P, Corbella J, Domingo JL, Etienne J-C, and Llobet JM, eds. *Metal Ions in*

Biology and Medicine, vol. 4, Proceedings of the Fourth International Symposium on Metal Ions in Biology and Medicine held in Barcelona (Catalonia) Spain, May 19-22, pp. 363-365 (1996).

Maiorino RM, Xu Z, and Aposhian HV. Determination and metabolism of dithiol chelating agents. XVII. In humans sodium 2,3-dimercaptopropane-1-sulfonate is bound to plasma albumin via disulfide formation and is found in the urine as cyclic polymeric disulfides. *J Pharmacol Exp Ther* 277:375-384 (1996).

Gonzalez-Ramirez D, Maiorino RM, Zuniga-Charles M, Xu Z, Hurlbut KM, Junco-Munoz P, Aposhian MM, Dart RC, Diaz GJH, Echeverria D, Woods JS, and Aposhian HV. Sodium 2,3-dimercaptopropane-1-sulfonate challenge test for mercury in humans: II. Urinary mercury, porphyrins and neurobehavioral changes of dental workers in Monterrey, Mexico. *J Pharmacol & Exp Ther* 272:1-11 (1995).

Aposhian HV. The Diversity of Arsenite Metabolism News Letter. Center for Toxicology, University of Arizona, Summer (1995)

Aposhian HV. Neurotoxicity of metals: the new challenges: Introductory remarks. *Toxicol* 97:1-2 (1995).

Aposhian HV, Gonzalez-Ramirez D, Maiorino RM, Zuniga-Charles M, Xu ZF, Hurlbut KM, Junco-Munoz P, Aposhian MM, and Dart RC. Mobilization of heavy metals by newer, therapeutically useful chelating agents. *Toxicol* 97:23-38 (1995).

Zakharyan R, Wu Y, Bogdan GM, and Aposhian HV. Enzymatic methylation of arsenic compounds: I. Assay and partial purification of arsenic methyltransferases from rabbit liver. *Chem Res Toxicol* 8:1029-1038 (1995).

Bogdan GM, Sampayo-Reyes A, and Aposhian HV. Arsenic binding proteins of mammalian systems: I. Isolation of three arsenite-binding proteins of rabbit liver. *Toxicol* 93:175-193 (1994).

Dart RC, Hurlbut KM, Maiorino RM, Mayersohn M, Aposhian HV, Boyer Hassen LV. Pharmacokinetics of meso-2,3-dimercaptosuccinic acid (DMSA) in lead poisoned patients and normal adults. *J Pediatr* 125:309-316 (1994).

Hurlbut KM, Maiorino RM, Mayersohn M, Dart RC, Bruce DC, and Aposhian HV. Determination and metabolism of dithiol chelating agents XVI. Pharmacokinetics of 2,3-Dimercapto-1-propanesulfonate after intravenous administration to human volunteers. *J Pharmacol Exp Ther* 268:662-668 (1994).

Maiorino RM, Aposhian MM, Xu Z, Li Y, Polt RL, and Aposhian HV. Determination and metabolism of dithiol chelating agents. XV. The meso-2,3-dimercaptosuccinic acid-cysteine (1:2) mixed disulfide, a major urinary metabolite of DMSA in the human, increases the urinary excretion of lead in the rat. *J Pharmacol Exp Ther* 267:1221-1226 (1993).

Aposhian HV, Levine DJ, Rivera M, and Fernando Q. Determination and metabolism of dithiol chelating agents: The zinc chelate of the dimethyl ester of meso-2,3-dimercaptosuccinic acid increases biliary excretion of cadmium and platinum. *Chem Res Toxicol* 6:208-214 (1993).

Atkins JM, Schroeder JA, Brower DL, and Aposhian HV. Evaluation of *Drosophila melanogaster* as an alternative animal for studying the neurotoxicity of heavy metals. *BioMetals* 5:111-120 (1992).

- Aposhian HV, Bruce DC, Alter W, Dart RC, Hurlbut KM, and Aposhian MM. Urinary mercury after administration of 2,3-dimercaptopropane-1-sulfonic acid: correlation with dental amalgam score. *FASEB J* 6:2472-2476 (1992).
- Zheng W, Perry DF, Nelson DL, and Aposhian HV. Choroid plexus protects cerebrospinal fluid against toxic metals. *FASEB J* 5:2188-2193 (1991).
- Maiorino RM, Dart RC, Carter DE, and Aposhian HV. Determination and metabolism of dithiol chelating agents. XII. Metabolism and pharmacokinetics of sodium 2,3-dimercaptopropane-1-sulfonate in humans. *J Pharmacol Exp Ther* 259:808-814 (1991).
- Aposhian HV, and Bruce DC. Binding of polonium-210 to liver metallothionein. *Radiat Res* 126:379-382 (1991).
- Rivera M, Bruck MA, Aposhian HV, and Fernando Q. The dimethyl ester of meso-2,3-dimercaptosuccinic acid and its interactions with Cd²⁺ and rabbit liver metallothionein I. *Chem Res Toxicol* 4:572-580 (1991).
- Rivera M, Levine DJ, Aposhian HV, and Fernando, Q. Synthesis and properties of the monomethyl ester of meso-dimercaptosuccinic acid and its chelates of lead(II), cadmium(II), and mercury(II). *Chem Res Toxicol* 4:107-114 (1991).
- Maiorino RM, Akins JM, Blaha K, Carter DE, and Aposhian HV. Determination and metabolism of dithiol chelating agents. X. In humans, meso-2,3-dimercaptosuccinic acid is bound to plasma protein via mixed disulfide formation. *J Pharmacol Exp Ther* 254:570-577 (1990).
- Bogdan GM, and Aposhian HV. N-(2,3-Dimercaptopropyl)phthalamidic acid (DMPA) increases polonium-210 excretion. *Biol Metals* 3:232-236 (1990).
- Zheng W, Maiorino RM, Brendel K, and Aposhian HV. Determination and metabolism of dithiol chelating agents. VII. The dithiol chelating agent DMPA increases biliary glutathione and biliary cadmium. *Fund Appl Toxicol* 14:598-607 (1990).
- Aposhian HV and Aposhian MM. Meso-2,3-dimercaptosuccinic acid. Chemical, pharmacological, and toxicological properties of an orally effective metal chelating agent. *Ann. Rev. Pharmacol. Toxicol* 30:279-306 (1990).
- Rivera M, Aposhian HV, and Fernando Q. Lead chelates of meso- and racemic dimercaptosuccinic acid. *J Inorg Biochem* 37:283-293 (1989).
- Aposhian HV and Aposhian MM. Newer developments in arsenic toxicity. *J Am Coll Toxicol* 8:1297-1305 (1989).
- Rivera M, Zheng W, Fernando Q, and Aposhian HV. Determination and metabolism of dithiol chelating agents. VIII. Metal complexes of meso-dimercaptosuccinic acid. *Toxicol Appl Pharmacol* 100:96-106 (1989).
- Maiorino RM, Bruce DC, and Aposhian HV. Determination and metabolism of dithiol chelating agents. VI. Isolation and identification of the mixed disulfides of meso-2,3-dimercaptosuccinic acid with 1-cysteine in human urine. *Toxicol Appl Pharmacol* 97:338-349 (1989).
- Aposhian HV, Maiorino RM, Dart RC, and Perry DF. Determination and metabolism of dithiol chelating agents. V. Urinary excretion of meso-dimercaptosuccinic acid in the human. *Clin Pharmacol Therap* 45:520-526 (1989).
- Maiorino RM, and Aposhian HV. Determination and metabolism of dithiol chelating agents. IV. Urinary excretion of meso-2,3-dimercaptosuccinic acid and

mercaptosuccinic acid in rabbits given meso-2,3-dimercaptosuccinic acid. *Biochem Pharmacol* 38:1147-1154 (1989).

Aposhian HV. Biochemical toxicology of arsenic. *Rev Biochem Toxicol* 10:265-299 (1989).

Aposhian MM, Aposhian HV, Domingo JL, Zheng W, and Dart RC. Radon decay products: DMPA decreases tissue polonium-210. *Plzen Lek Sborn Suppl*. 1988:99-100 (1988).

Maiorino RM, Weber GL, and Aposhian HV. Determination and metabolism of dithiol chelating agents. III. Formation of oxidized metabolites of 2,3-dimercaptopropane-1-sulfonic acid in rabbit. *Drug Metab Dispos* 16:455-463 (1988).

Maiorino RM, Barry TJ, and Aposhian HV. Determination and metabolism of dithiol chelating agents: Electrolytic and chemical reduction of oxidized dithiols in urine. *Anal Biochem* 160:217-226 (1987).

Aposhian HV, Dart RC, Aposhian MM, and Dawson BV. Tissue decorporation of polonium-210 in rats by DMPA. *Res Comm Chem Pathol Pharmacol* 58:157-171 (1987).

Aposhian HV, Maiorino RM, Aposhian MM, Dart RC, Tobias PS, Zheng W, and Perry DF. Dimercapto metal binding agents: decorporation of Po-210 and Cd-109 in the rat and metabolic studies in the human. *Environmental Health Series-20*. World Health Organization: Denmark, pp. 215-218 (1987).

Maiorino RM, Weber GL, and Aposhian HV. Fluorometric determination of 2,3-dimercaptopropane-1-sulfonic acid and other dithiols by precolumn derivatization with bromobimane and liquid chromatography. *J Chromatog (Biomed.)* 374:297-310 (1986).

Maiorino RM and Aposhian HV. Dimercaptan metal-binding agents influence the biotransformation of arsenite in the rabbit. *Toxicol Appl Pharmacol* 77:240-250 (1985).

Aposhian HV, Maiorino RM, Aposhian MM, Hsu CA, and Stine ER. Water soluble dithiol metal binding agents -- Efficacies and additional modes of action. *Plzen Lek Sborn Suppl* 49:47-51 (1985).

Stine ER, Hsu CA, Hoover TD, Aposhian HV, and Carter DE. N-(2,3-dimercaptopropyl) phthalamidic acid: Protection, in vivo and in vitro, against arsenic intoxication. *Toxicol Appl Pharmacol* 75:329-336 (1984).

Aposhian HV, Carter DE, Hoover TD, Hsu CA, Maiorino RM, and Stine E. EMSA, DMPS, and DMPA -- as arsenic antidotes. *Fund Appl Toxicol* 4:S58-S78 (1984).

Putney S, Herlihy W, Royal N, Pang H, Aposhian HV, Pickering L, Belagaje R, Biemann K, Page D, Kuby S, and Schimmel P. Rabbit muscle creatine phosphokinase. *J Biol Chem* 259:14317-14320 (1984).

Slilaty SN and Aposhian HV. Gene transfer by polyoma-like particles assembled in a cell-free system. *Science* 220:725-727 (1983).

Hoover TD and Aposhian HV. BAL increase the 74As content of rabbit brain. *Toxicol Appl Pharmacol* 70:160-162 (1983).

Aposhian HV, Hsu CA, and Hoover TD. DL- and meso-dimercaptosuccinic acid: In vitro and in vivo studies with sodium arsenite. *Toxicol Appl Pharmacol* 69:206-213 (1983).

- Hsu CA, Aposhian HV, Heydolph S, and Parr W. Optical isomers of 2,3-dimercapto-1-propanesulfonate: Antidotal activity, in vitro and in vivo, against sodium arsenite. *J Pharmacol Exp Ther* 224:314-318 (1983).
- Aposhian HV. DMSA and DMPS -- water soluble antidotes for heavy metal poisoning. *Ann Rev Pharmacol Toxicol* 23:193-215 (1983).
- Aposhian HV, Mershon MM, Brinkley FB, Hsu CA, and Hackley BE. Anti-lewisite activity and stability of meso-dimercapto acid and 2,3-dimercapto-1-propanesulfonic acid. *Life Sci* 31:2149-2156 (1982).
- Slilaty SN, Berns KI, and Aposhian HV. Polyoma-like particle: Characterization of the DNA encapsidated in vitro by polyoma empty capsids. *J Biol Chem* 257:6571-6575 (1982).
- Aposhian HV. Meso-dimercaptosuccinic acid and 2,3-dimercapto-1-propanesulfonic acid: Effective water soluble analogs of British anti-lewisite. *Adv Enzyme Reg* 20:301-319 (1982).
- Aposhian HV, Tadlock CH, and Moon TE. Protection of mice against the lethal effects of sodium arsenite -- a quantitative comparison of a number of chelating agents. *Toxicol Appl Pharmacol* 61:385-392 (1981).
- Tadlock CH and Aposhian HV. Protection of mice against the lethal effects of sodium arsenite by 2,3-dimercapto-1-propanesulfonic acid and dimercaptosuccinic acid. *Biochem Biophys Res Comm* 94:501-507 (1980).
- Barr SM, Keck K, and Aposhian HV. Cell-free assembly of a pancreatic DNase-resistant and salt-resistant polyoma-like particle from empty capsids and DNA. *Virology* 96:656-659 (1979).
- Aposhian HV and Zakharyan RA. Assembly of a polyoma-like particle from empty capsids and DNA in a cell-free system. *Adv Enzyme Reg* 18:275-287 (1979).
- Aposhian HV, Barr SM, and Keck, K. Experimental gene delivery system for mammalian cells -- the polyoma pseudovirus system. *Adv Enzyme Reg* 16:275-288 (1978).
- Aposhian HV. Pseudovirions in animals, plants and bacteria. In: *Comprehensive Virology* (Fraenkel-Conrat H and Wagner RR, eds.), Vol. 5. Plenum Publishing Co., pp. 155-218 (1975).
- Aposhian HV, Thayer RE, and Qasba PK. Formation of nucleo-protein complexes between polyoma empty capsids and DNA. *J Virol* 15:645 (1975).
- Kashmiri SVS and Aposhian HV. Degradation of pseudoviral DNA after infection of mouse cells with polyoma pseudovirions. *Proc Natl Acad Sci USA* 71:3834 (1974).
- Yelton DB and Aposhian HV. Polyoma pseudovirions. II. Influence of host cell on pseudovirus production. *J Virol* 12:1065 (1973).
- Aposhian HV, Qasba PK, Yelton DB, Pletsch QA, and Sethi VS. The polyoma pseudovirion -- Its properties, production and use in transferring DNA to mouse and human cells. In: *Cellular Modification and Genetic Transformation by Exogenous Nucleic Acids* (Beers RF and Tilghman RC, eds.). The Johns Hopkins University: Baltimore (1973).
- Aposhian HV. A review of mammalian gene transfer systems and their future implications. *Inter Medicos* 15:129 (1972).
- Yelton DB and Aposhian HV. Polyoma pseudovirions. I. Sequence of events in primary mouse embryo cells leading to pseudovirus production. *J Virol* 10, 340 (1972).

Aposhian HV. Gene therapy, gene therapy. In: Graduate School Chronicle, Vol. V, No. 2. University of Maryland, p. 7 (1972).

Qasba PK, Yelton DB, Pletsch QA, and Aposhian HV. Properties of polyoma pseudovirions in mouse and human cells. In: Molecular Studies in Viral Neoplasia, A Collection of Papers Presented at the 25th Annual Symposium on Fundamental Cancer Research. Williams and Wilkins Co.: Baltimore, p. 169 (1972).

Aposhian HV, Qasba PK, Osterman JV, and Waddell A. Polyoma Pseudovirions: An experimental model for the development of DNA for gene therapy. Fed Proc 13:1310 (1972).

Aposhian HV, Qasba PK, Osterman JV, and Waddell A. Prospects for designed genetic change. A report from the National Advisory General Medical Sciences Council. DHEW Publication No. (NIH) 72-78 (1972).

Qasba PK and Aposhian HV. DNA and gene therapy: Transfer of mouse DNA to human mouse embryonic cells by polyoma pseudovirions. Proc Natl Acad Sci USA 68:2345 (1971).

Osterman JV, Waddell A, and Aposhian HV. Gene therapy systems: The need, experimental approach and implications. In: Drug Metabolism in Man. NY Acad Sci 179:514 (1971).

Aposhian HV. Penicillamine and analogous chelating agents. In: Drug Metabolism in Man. NY Acad Sci 179:481 (1971).

Osterman JV, Waddell A, and Aposhian HV. DNA and gene therapy. Uncoating of polyoma pseudovirus in mouse embryo cells. Proc Natl Acad Sci USA 67:37 (1970).

Aposhian HV. The use of DNA for gene therapy -- The need, experimental approach and implications. Perspect Bio Med 14:98 (1970).

Koh JK, Waddell AD, and Aposhian HV. The synthesis and breakdown of nucleic acids in mammalian cells transformed by oncogenic viruses. I. Purification and properties of an endonuclease from baby hamster kidney cells transformed by polyoma virus. J Biol Chem 245:4698 (1970).

Aposhian HV, Friedman N, Nishihara M, Heimer EP, and Nussbaum AL. Sequential cleavage of dinucleotides from DNA by SP3 DNase III. Substrate specificity and initiation of the hydrolysis from the 5'-terminus of polynucleotides. J Mol Biol 49:367 (1970).

Nishihara M, Friedman N, and Aposhian HV. Biological activity of 5'-hydroxymethyluracil and its deoxynucleoside in non-infected and phage-infected *Bacillus subtilis*. J Virol 3:164 (1970).

Trilling DM and Aposhian HV. Sequential cleavage of dinucleotides from DNA by phage SP3 DNase. Proc Natl Acad Sci USA 60:214 (1968).

Aposhian HV. Biosynthesis of the components of DNA phages. In: Molecular Basis of Virology (H. Fraenkel-Conrat, ed.). Reinhold Book Corp. (1968).

Nishihara M, Chrambach A, and Aposhian HV. The deoxycytidylate deaminase found in *Bacillus subtilis* infected with phage SP8. Biochem 6:1877 (1967).

Aposhian HV and Tremblay GY. Deoxythymidylate-5'-nucleotidase purification and properties of an enzyme found after infection of *Bacillus subtilis* with phage SPC5. J Biol Chem 239:222 (1964).

Richardson CC, Schildkraut CL, Aposhian HV, and Kornberg A. Further purification and properties of DNA polymerase of *E. coli*. J Biol Chem 239:222 (1964).

- Richardson CC, Schildkraut CL, Aposhian HV, Kornberg A, Bodmer W, and Lederberg J. Studies on the replication of DNA by E. coli polymerase. In: Information Macromolecules (H.J. Vogel, V. Bryson, and J.O. Lampen, eds.). Academic Press: New York, p. 13 (1963).
- Aposhian HV, and Kornberg A. The polymerase formed after T2 bacteriophage infection of E. coli: A new enzyme. *J Biol Chem* 237:519 (1962).
- Aposhian HV. Biochemical and pharmacological properties of the metal binding penicillamine in Wilson's disease. (Walshe JM and Cumings JN, eds.). Blackwell Scientific Publishers, Ltd. (1961).
- Aposhian HV. Biochemical and pharmacological properties of the metal binding agent penicillamine. *Fed Proc* 20:195 (1961).
- Lambooy JP and Aposhian HV. The biological activity of 6-ethyl-9-(1'-D-ribityl) isoloazazine. *J Nutrition* 71:182 (1960).
- Aposhian HV, Pointer NS, and Aposhian MM. Reversal of the inhibitory activity of chlorpromazine by calcium. *Proc Soc Exp Biol Med* 100:512 (1959).
- Aposhian HV, Blair RM, Morris M, and Smithson CH. The reversal of the inhibitory activity of L-penicillamine by branched chain amino acids. *Biochem Biophys Acta* 36: 93 (1959).
- Aposhian HV and Aposhian MM. N-acetyl-DL-penicillamine. A new oral protective agent against the lethal effects of mercuric chloride. *J Pharmacol Exp Ther* 126:131 (1959).
- Blair RM and Aposhian HV. The inhibition of E. coli by L-penicillamine and its reversal by isoleucine, valine or leucine. *Biochem Biophys Acta* 30:214 (1958).
- Aposhian HV. Protection by D-penicillamine against the lethal effects of mercuric chloride. *Science* 128:93 (1958).
- Aposhian HV. Penicillamine and dimercaprol. *Lancet* 859 (1958).
- Aposhian HV, Wolff SM, and Rhea WG, Jr. The inhibition of serine metabolism in *Leuconostoc mesenteroids* P-60. I. Microbiological activity of D- and L-penicillamine. *Arch Biochem Biophys* 71:442 (1957).
- Aposhian HV and Lambooy JP. The in vivo synthesis of diethylriboflavin phosphate. *J Am Chem Soc* 77:6368 (1955).
- Aposhian HV and Lambooy JP. The synthesis of 6-ethyl-9-(D-1'ribityl)-isoaloxazine. *J Am Chem Soc* 76:1307 (1954).
- Lambooy JP and Aposhian HV. The biological activity of diethyl riboflavin. *J Nutrition* 47:539 (1952).
- Kochakian CD and Aposhian HV. The in vitro metabolism of 3-17-androstanediol of liver and kidney. *Arch Biochem Biophys* 37:442 (1952).
- Aposhian HV and Lambooy JP. Retardation of growth of Walker rat carcinoma 256 by administration of diethyl riboflavin. *Proc Soc Exp Biol Med* 78:197 (1951).

CERTIFICATE OF SERVICE

I hereby certify that on August 31, 2007, I served the foregoing **Expert Report from Dr. H. Vasken Aposhian, PhD** on the following individual(s):

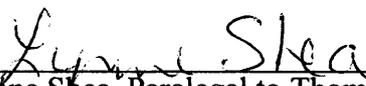
John Fabry, Esq.
Williams Kherkher Hart Boundas, LLP
8441 Gulf Freeway, Suite 600
Houston, TX 77017-5001

Vincent Matanoski, Esq.
Mark Raby, Esq.
US Department of Justice
Torts Branch, Civil Division
1425 New York Avenue NW
Suite 3100
Washington DC, 20005

By United Parcel Service, next day delivery.

Petitioners specifically authorize the Court and the Office of Special Masters to post this document, and any attachments or exhibits thereto, on the Court/OSM website, expressly waiving any confidentiality as to the contents of these materials. Petitioners expressly wish to publicly disclose this filing in any other forum designated by the Court or the OSM.

WILLIAMS LOVE O'LEARY & POWERS, P.C.



Lynne Shea, Paralegal to Thomas B. Powers
Of Attorneys for Petitioners' Steering Committee

10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
66
67
68
69
70
71
72
73
74
75
76
77
78
79
80
81
82
83
84
85
86
87
88
89
90
91
92
93
94
95
96
97
98
99
100